No. 2

Vol. 33 June 2008 -

### CONTENTS

	Page
Foraging frequency and pattern of movement of different Apis species on parental lines of Brassica napus L.: Jasvir Singh.	91
Time of adult emergence and sex ratio of rice blue beetle, <i>Leptispa pygamaea</i> Baly (Coleoptera: Chrysomelidae) and extent of damage caused by the pest to two	,
rice varieties: K. Karthikeyan, Sosamma Jacob.	101
Histomorphological derangements in the ovary of <i>Oryctes rhinoceros</i> (Coleoptera: Scarabaeidae) treated with methanolic extract of <i>Annona squamosa</i> leaves:	
K. B. Sreelatha, P. R. Geetha	107
tivars: Wajid Hasan, M. S. Ansari, Haidar Ali	113
New reports of predatory mites (Acari: Prostigmata, Mesostigmata) from medicinal plants of Darjeeling district, West Bengal, India with description of a new	
species: Indranil Roy, Salil K. Gupta, Goutam K. Saha	119
Jyothi Tilak.	129
SHORT COMMUNICATIONS	
Comparative genotoxicity of alpha-cyano pyrethroids on <i>Drosophila melanogaster</i> : Namrata Rana, Nishi Saxena, Harendra N. Sharma, P. N. Saxena.	135
Effect of age of the host plant on the performance of <i>Ceutorhynchus portulacae</i> Marshal (Coleoptera: Curculionidae), a herbivore of <i>Portulaca oleracea</i> : P. N. Ganga	
Visalakshy.	139
Butea monosperma (Lam.), a new host of Lampides boeticus Linnaeus in Rajastan, India: Kan Singh, R. Swaminathan.	143
Identification of promising multivoltine × bivoltine hybrids of the mulberry silk-	147
worm, Bombyx mort L.: R. Nirupama, Ravindra Singh, D. Gangopadhyay Effect of feeding larvae of Helicoverpa armigera (Hubner) on Chickpea (Cicer ariet-	14/
inum L.) treated with chemical and organic fertilizers: B. K. Singh, R. P. Singh.	151
A new host plant record for <i>Oberea artocarpi</i> Gardner (Coleoptera: Cerambycidae):	157
K. D. Prathapan	131



### **ENTOMON**

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### Foraging frequency and pattern of movement of different *Apis* species on parental lines of *Brassica* napus L.

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ABSTRACT: Study conducted on number of flowers visited/min and pattern of movement of different *Apis* species on parental lines of *Brassica napus* to evaluate their pollination efficiency, revealed that various bee species according to their foraging rate were: *A. cerana* = *A. mellifera* > *A. dorsata* > *A. florea*. Foraging frequency of *Apis* species was found to be maximum at 1200 h, followed by 1000, 1400 and 1600 hours. Number of flowers visited/min by all the species of honeybees was significantly more on cytoplasmically male sterile line as compared to that on restorer line. The effective intersexual flower visits (visits from R to CMS lines) of bees were extremely low (3.300–3.800%) which could not be considered satisfactory for hybrid seed production. All the four bee species showed very high percentage of intrasexual flower visits, however these bee species did not differ significantly for all types of intersexual and intrasexual flower visits. Bees tend to remain on the same parental line on which they started foraging, thus indicating higher floral constancy in their foraging behaviour. Effective visits were slightly more in 2:4 row ratio plots followed by 1:4 and 1:8 row ratios. © 2008 Association for Advancement of Entomology

KEYWORDS: Apis cerana, Apis dorsata, Apis florea. Apis mellifera, Brassica napus, pollination efficiency, hybrid seed production

### INTRODUCTION

Honeybees are the most efficient and abundant amongst insect pollinators of *Brassica* Crops. In general, foraging rate (number of flowers visited/min) and pattern of movement of different bee species on parental lines of *B. napus* determine their pollination efficiency. More the foraging rate of a bee, more are the chances of pollination. Foraging rate of any insect visitor depends on many factors like instinctive foraging behaviour. length of proboscis (Inouye, 1980). floral structure (Free, 1970), particularly the corolla depth (Gilbert, 1980), type and quantity of floral rewards (Rao and Suryanarayana, 1990; Rao, 1991; Dashad *et al.*, 1992). On small sized flowers

92 J. Singh

foraging speed of bees is faster (Mohr and Jay, 1988). Hours of day also affect the foraging frequency of insects.

Honeybees being potential agents of pollen transfer are effectively used for pollination and production of hybrid seeds. The flowers on cytoplasmically male sterile (cms) line must be visited by nector seeking pollen coated insects that have earlier visited the flowers of restorer (R) line during the same visit. Intersexual flower visits of honeybees may not take place to a desired extent because flowers of both the parental lines are not equally attractive to them and they show higher degree of floral constancy.

Present study is an attempt to compare foraging rates of different *Apis* species and their pattern of movement between R and CMS lines in hybrid seed production plots of *B. napus* to assess their pollinating efficiency.

### MATERIALS AND METHODS

Experimental plots for hybrid seed production were raised under Punjab conditions by sowing seeds of cytoplasmically male sterile (CMS) line (TCMS-PR-05) and restorer (R) line (TFR-91) of *B. napus* hybrid PGSH-51, by following recommendations of Punjab Agricultural University, Ludhiana (Punjab) during years 2003–2004 and 2004–2005. Selected male to female row ratios were 2:4, 1:4 and 1:8, the recommended ratio being 2:4. Early flowering R line plants were detopped to make the flowering of two lines synchronous. Two colonies each of *A. mellifera*, *A. cerana*, and *A. florea* (by comb transfer technique) were placed in the field. *A. dorsata* hives were present in nearby trees. Observations in each experiment were taken at 1000, 1200, 1400 and 1600 hours. Following two experiments were conducted.

For foraging frequency assessment of *Apis* species on parental lines of *B. napus* the number of flowers visited/min by a particular bee species on male and female lines, was counted with help of stop clock at selected hours during day. This experiment was conducted on 20 alternating days in full blooming season of crop during year 2004 and 2005. The pooled mean of two observations has been considered.

Pattern of movement of *Apis* species between parental lines of *B. napus* was studied in plots of various row ratios by following a particular bee for one minute at selected timing of the day and the percentage of male to female, male to male, female to female and female to male visits were recorded. Observations were started from male and female rows alternatingly and recorded separately. This experiment was conducted for 20 alternating days in full flowering season of crop during year 2005.

Data collected from above two experiments were consolidated, tabulated, transformed ( $\sqrt{x} + 1$  transformations and arc sin transformations in experiment no. 1 and 2 respectively) and subjected to analysis of variance. Significance was tested at 5 per cent level.

### RESULTS AND DISCUSSION

Foraging rates of different honeybee species at the selected hours of day on R and CMS lines are shown in Table 1A. On the basis of mean number of flowers visited/min (Table 1B) various bee species can be arranged as: A. cerana (19.143 flowers/min) = A. mellifera (18.836 flowers/min) > A. dorsata (14.460 flowers/min) > A. florea (7.623 flowers/min). The results of this study are in line with the observations recorded by Desh Raj and Rana (1994) who found that foraging speed of A. cerana was non significantly more than that of A. mellifera on rapeseed (B. campestris var. brown sarson) bloom. Observations were recorded on foraging speed of these two bee species on apple bloom (Verma and Dulta, 1986) and on plum flowers (Rana, 1989). Jhajj et al. (1996) found the foraging speed of bee species as: A. mellifera > A. dorsata > A. florea on raya and brown sarson.

Various day hours according to foraging rate of *Apis* species (Table 1C) could be arranged as: 1200 h (16.359 flowers/min) > 1000 h (15.166 flowers/min) > 1400 h (14.792 flowers/min) > 1600 h (13.744 flowers/min). Highest foraging rate was observed at 1200–1300 hours on *Brassica* crops by many workers (Dhaliwal and Bhalla, 1980; Desh Raj and Rana, 1994; Anonymous, 1999). Comparatively more foraging rate of honeybees at 1200 h might be due to the availability of floral rewards declined by this time due to increased foraging activity of bees as observed by Meyerhoff (1954) in case of swede rape. So, less time was needed to take forage from a flower and more number of flowers were visited by a bee to collect a required load of pollen or nectar. Hours of the day played an important role in regulating foraging rate of bees.

Data of present study (Table 1D) showed that average foraging rates of bees (Irrespective of bee species and day hours) was significantly higher on CMS line (16.647 flowers/min) than that on R line (13.384 flowers/min). These results are similar to those of Dashad et al. (1992), who observed that foraging rates of insect visitors were different on different cultivars of apple. Difference in foraging rates of Apis species on R and CMS lines may be due to the difference in shape and size of flowers, expansion of their petals, type and quantity of floral rewards (Rao and Suryanarayana, 1990; Rao, 1991; Dashad et al., 1992) and some other morphological and physiological traits of flowers of two lines. The R line flowers offer both nectar and pollen, hence the bees visiting them were pollen and/or nectar gatherers. They spent longer time on individual flower and thus have lower foraging rate. While flowers of CMS line offer only the nectar as they do not produce pollen, the foraging bees were only nectar gatherers. Therefore, honeybees spent less time on CMS line flowers and have higher foraging rate. Adegas and Nogueira (1992) also observed that nectar collecting bees spent less time per flower compared to pollen gatherers on B. napus L. var *oleifera* (cultivar CTC-4).

Data in Table 2, 3 showed that in plots of all the row ratios, either the observations were started from male or female line, intersexual flower movement, especially male to female flower visits of bees (effective for cross pollination) were extremely low. Comparatively more of such movements were recorded in case of *A. florea* (1.375–

TABLE 1A. Mean number of flowers visited/min by different Apis species on parental lines of B. napus at selected hours of day

Day hours	Apis species	Number of f	lowers visited/min
		R line	CMS Line
1000 h.	A. mellifera	16.583	20.644
		(4.1860)	(4.6520)
	A. cerana	18.063	20.428
		(4.3589)	(4.6273)
	A. dorsata	12.416	16.750
		(3.6618)	(4.2130)
	A. florea	7.000	9.442
	-	(2.8262)	(3.2297)
1200 h.	A. mellifera	18.756	23.146
		(4.4377)	(4.9102)
	A. cerana	18.146	23.213
		(4.3668)	(4.9167)
	A. dorsata	14.338	16.354
		(3.9156)	(4.1501)
	A. florea	7.400	9.519
		(2.8917)	(3.2413)
1400 h.	A. mellifera	16.190	19.906
		(4.1411)	(4.5578)
	A. cerana	16.250	22.575
		(4.1532)	(4.8413)
	A. dorsata	13.693	14.500
		(3.8300)	(3.9361)
	A. florea	5.814	9.406
		(2.6083)	(3.2122)
1600 h.	A. mellifera	16.022	19.438
		(4.1256)	(4.5203)
	A. cerana	15.250	19.219
		(4.0304)	(4.4893)
	A. dorsata	13.750	13.875
		(3.8398)	(3.8559)
	A. florea	4.464	7.938
		(2.3372)	(2.9833)

TABLE 1B. Foraging rates of different *Apis* species irrespective of day hours and parental lines

Apis species	A. mellifera	A. cerana	A. dorsata	A. florea
Number of flowers	18.836	19.143	14.460	7.623
visited/min.	(4.4413)	(4.4730)	(3.9253)	(2.9162)

TABLE 1C. Foraging rates of *Apis* species at different day hours, irrespective of bee species and parental lines

Day hour	1000 h	1200 h	1400 h	1600 h
Number of flowers visited/min.	15.166	16.359 (4.1037)	14.792 (3.9100)	13.744

TABLE 1D. Foraging rates of *Apis* species on R and CMS rows irrespective of bee species and day hours

Parental line	R Line	CMS Line
Number of flower visited/min.	13.384	16.647
	(3.7319)	(4.1460)

CD0.05

For day hours 0.102

For Apis spp. 0.102

For Parental lines 0.072

For interaction day hours x Apis spp. Non significant

For interaction day hours x Parental lines Non Significant

For interaction Apis spp. X Parental lines 0.145

For interaction day hours x Apis spp. X Parental line. Non significant.

Figures in parentheses are  $\sqrt{n+1}$  transformations.

3.375%), followed by A. mellifera (0.775–2.700%) and A. cerana (0.425–1.105%). Minimum percentage of such visits was recorded in case of A. dorsata (0.300–0.962%). However all the bee species did not differ significantly for different types of inter-sexual or intra-sexual flower visits. Comparatively more percentage of male to female flower visits of A. florea bees may be due to fact that this bee species is a dominant nectar gatherer and both male and female lines provide nectar. More nectar gathering behaviour of A. florea was observed by Panda et al. (1995) on niger bloom.

Intra-sexual flower visits were much higher than inter-sexual flower visits in case of all species of *Apis*. When observations were started from male line (Table 2, the male to male flower visits were dominant (76.900–95.475%) followed by female to female (3.062–18.970%) Similarly, when observations were started from female line (Table 3, the female to female flower visits were maximum (69.075–93.308%) followed by male to male visits (5.245–23.750%). In both the conditions, inter-sexual flower movement was very low with a range of 0.300–3.800. Bees tried to remain on same parental line on which they started foraging, indicating thereby, a clear cut floral constancy in their foraging behaviour. A minor percentage of bees showed crossing over between male and female lines bring about low degree of cross pollination. Similar type of results regarding low percentage of inter-sexual flower visits and higher percentage of intra-sexual flower movement of *A. mellifera* in hybrid seed production plots of B. *napus* have also been reported earlier (Anonymous, 1999). Honeybees tend to visit within an inbred rather than between lines (Johnson, 1972). Faulkner (1974) also observed

96 J. Singh

TABLE 2. Movement of *Apis* species between male and female lines of *B. napus* (while starting observation from male row)

Male to female Row ratio			types of vis	its of <i>Apis</i> s	pecies
Itow Iulio	Apis species	M-F	M-M	F-F	F-M
2:4	A. mellifera	1.347	92.435	5.010	1.208
		(6.432)	(74.276)	(12.785)	(6.112)
	A. cerana	0.862	93.975	4.300	0.863
		(5.108)	(75.902)	(11.905)	(5.108)
	A. dorsata	0.738	95.475	3.062	0.725
		(4.537)	(78.000)	(9.951)	(4.504)
	A. florea	2.995	80.880	13.000	3.125
	y .	(9.821)	(64.118)	(21.107)	(10.011
	Average	1.486	90.691	6.343	1.480
	C	(6.474)	(73.074)	(13.937)	(6.434)
	CD 0.05 for A	pis spp. N			
	for movement	1.527			
	For interactio	n <i>Apis</i> spp.	x movemer	ats = 3.054	
1:4	A. mellifera	1.050	87.237	10.825	0.888
	,	(5.700)	(69.177)	(19.144)	(5.035)
	A. cerana	0.525	90.127	8.898	0.450
		(4.141)	(71.819)	(17.212)	(3.840)
	A. dorsata	0.523	91.282	7.692	0.500
		(3.882)	(73.006)	(15.973)	(3.804
	A. florea	2.350	76.900	18.375	2.375
	,	(8.660)	(61.320)	(25.360)	(8.703
	Average	1.112	86.386	11.448	1.053
	C	(5.596)	(68.830)	(19.422)	(5.346
	CD 0.05 for A	<i>pis</i> specie		icant	,
	for movement	. ,			
	for interaction		x movement	ts 2.980	
1:8	A. mellifera	0.775	79.455	18.970	0.688
	3	(4.876)	(63.102)	(25.780)	(4.616)
	A. cerana	0.425	85.975	13.150	0.450
		(3.719)	(68.192)	(21.073)	(3.816
	A. dorsata	0.400	86.722	12.578	0.300
		(3.390)	(68.696)	(20.725)	(2.933
	A. florea	1.375	87.825	9.550	1.250
	Ţ.	(6.661)	(69.616)	(17.978)	(6.337
	Average	0.744	84.994	13.562	0.672
	C	(4.662)	(67.401)	(21.389)	(4.426
	CD 0.05 for A				,
	for movement				
	for interaction		x movemen	its 2.727	

Figures in parentheses are arc sine transformations.

TABLE 3. Movement of *Apis* species between male and female lines of *B. napus* (while starting observations from female row)

Male to female row ratio	Percer		ous types of	visits of Apa	is
	Apis species	M-F	M-M	F-F	F-M
2:4	A. mellifera	2.700	10.608	83.467	3.225
		(9.170)	(18.748)	(66.348)	(9.969)
	A. cerana	1.105	13.970	83.600	1.325
		(5.930)	(21.805)	(66.278)	(6.459)
	A. dorsata	0.962	12.072	85.965	1.000
		(5.328)	(20.249)	(68.116)	(5.438)
	A. florea	3.375	23.750	69.075	3.800
	,	(10.452)	(29.154)	(56.222.)	(11.156)
	Average	2.036	15.100	80.527	2.338
		(7.720)	(22.489)	(64.241)	(8.255)
	CD 0.05 for A for movement for interaction	l <i>pis</i> spp. noi ts 1.968	n significant		, ,
1.4	4 11:0	2.060	0.530	06.017	2.450
1:4	A. mellifera	2.060	8.538	86.917	2.458
	4	(7.946)	(16.805)	(69.071)	(8.676)
	A. cerana	1.060	9.590	88.100	1.250
		(5.896)	(17.872)	(69.957)	(6.391)
	A. dorsata	0.700	7.450	90.950	0.900
	_	(4.594)	(15.749)	(72.641)	(5.217)
	A. florea	2.900	19.900	74.225	2.975
		(9.671)	(26.470)	(59.509)	(9.803)
	Average	1.680	11.370	85.048	1.902
		(7.027)	(19.224)	(67.795)	(7.522)
	CD 0.05 for A	A <i>pis</i> spp. noi	n significant		
	for movemen				
	for interaction	n <i>Apis</i> spp. x	movement	3.456	
1:8	A. mellifera	1.525	7.218	89.782	1.475
		(6.843)	(15.267)	(71.694)	(6.730)
	A. cerana	0.925	5.245	92.855	0.975
		(5.456)	(13.052)	(74.682)	(5.593)
	A. dorsata	0.300	6.167	93.308	0.225
		(3.037)	(14.257)	(75.141)	(2.641)
	A. florea	1.600	8.525	88.275	1.600
	-	(7.204)	(16.957)	(70.013)	(7.204)
	Average	1.088	6.789	91.055	1.069
		(5.635)	(14.883)	(72.833)	(5.542)
	CD 0.05 for A	. ,		(.2.000)	(2.2.2)
	for movement		. s.giouiit		
	for interaction		movement	3 193	
	701 Interaction			, 5.175	

98 J. Singh

that a mean ratio of 30:1 was present between visits of bees to flowers on the same inbred and those flowers on different inbreds in case of cauliflowers while such ratio was recorded to be 33:1 in case of Brussels sprout (Faulkner, 1976). Similar type of conclusions were drawn regarding effective bee visits in hybrid seed production plots of Brussels sprout (Free and Williams, 1983) and cotton (Eisikowitch and Loper, 1984).

Low percentage of inter-sexual flower visits was due to the fact that male flowers of *B. napus* were more attractive to bees as compared to the CMS flowers due to their larger size, better looks and availability of both pollen and nectar. Since there is no pollen for bees to collect from male sterile flowers, they visit the flowers to collect nectar only. Flowers of CMS line compete poorly for attention of pollinators. More attractiveness of male line flowers of *B. napus* has also been confirmed by many workers (Mesquida and Renard, 1979a; Ohsawa and Nawal, 1988; Anonymous, 1999). Difference in morphology, physiology, floral rewards, foraging cues, floral events, aroma chemistry etc of male and female flowers causes selective foraging by honey bees in case of B. napus (Renard and Mesquida, 1987; Mosquida *et al.*, 1987; Ohsawa and Nawal, 1988; Anonymous, 1999).

Minor increase in intersexual flower visits of bees was found with changing row ratios from 1:8, through 1:4 to 2:4, either observations were started from male or female line. This is due to the fact that frequency of male rows was higher in case of 2:4 row ratio which is recommended ratio, followed by 1:4 and 1:8 planting ratio.

Any species of *Apis* which has higher foraging rate is considered to be an efficient pollinator as it may pollinate more number of flower/min. Thus *A. cerana* and *A. mellifera* were found better pollinators and they were followed by *A. dorsata* and *A. florea*. Hours of the day also played an important role and maximum foraging rate was recorded at 1200 h. Significantly more number of flowers were visited/min by bees on CMS line as compared to that on R line.

Effective intersexual flower visits (movement from male to female line) of all the four *Apis* species were low and could not be regarded as satisfactory for hybrid seed production. Therefore research should be oriented to select/breed lines which are equally attractive to honeybees, have synchronized flowering and provide floral rewards of equal value. Essential factors for bee attractiveness should be studied and incorporated into new hybrid system. Recommended planting ratio of 2:4 was found suitable as far as pollination causing visits were concerned.

### REFERENCES

Adegas, J. E. B. and Nogueira, C. R. H (1992) Entomophilous pollination in rape (*Brassica napus* L. var. *oleifera*) in Brazil. *Apidologie* **23**: 203–209.

Anonymous, (1999) *Annual Report*, All India co-ordinated project on honeybee Res. and Trg. C.C.S, HAU Hissar, 16–20.

Dashad, S. S., Sharma, J. K. and Yadav, P. R. (1992) Foraging speed of insect visitors to the blooms of various cultivars of apple (*Malus domestica Borkh*). *Indian Bee J.* **54**: 92–98.

Desh, Raj and Rana, V. K. (1994) Time spent by *Apis mellifera* L. and *Apis cerana indica* F. foragers on rapeseed bloom. *J. Ent. Res.* **18**: 335–339.

- Dhaliwal, H. S. and Bhalla, O. P. (1980) The foraging ecology of *Apis cerana indica*. In: *Proc. 2nd Internat. Conf. Apic. Trop. Clim.*, New Delhi, 513–527.
- Eisikowitch, D. and Loper, G. M. (1984) Some aspects of flower biology and bee activity on hybrid cotton in Arizona, U.S.A. J. Apic. Res. 23: 243–248.
- Faulkner, G. J. (1974) Factors effecting fieldscale production of seed of f<sub>1</sub> hybrid Brussels sprout. *Ann. Appl. Biol.* 77: 181–190.
- Faulkner, G. J. (1976) Honeybee behaviour effected by plant height and flower colour in Brussels sprouts. *J. Apic. Res.* **15**: 15–18.
- Free, J. B. (1970) Insect Pollination of Crop Plants, Academic Press, London and New York.
- Free, J. B. and Williams, I. H. (1983) Foraging behaviour of honeybees and bumble bees on Brussels sprout grown to produce hybrid seed. *J. Apic. Res.* 22: 94–97.
- Gilbert, F. S. (1980) Flower visiting by hoverflies (*Syrphidae*). *Ann. Rev. Ecol. Syst.* **6**: 139–170. Inouye, (1980) The effect of proboscis and corolla tube lengths on patterns and rates of flower visitation by bumble bees. *Oecologia* **45**: 197–201.
- Jhajj, H. S., Gatoria, G. S. and Bakhetia, D. R. C. (1996) Three Decades of Bee Keeping Research in the Punjab, Nat. Agric. Tech. inf. Centre, Ludhiana (Pb.), 23–27.
- Johnson, A. G. (1972) Problems in breeding and seed production of hybrid sprout. Commercial Grower 4013: 787–789.
- Mesquida, J. and Renard, J. (1979a) Entomorphilous pollination of male sterile strains of winter rapeseed (Brassica napus L. Metzger Var. oleifera) and a preliminary study of alternating devices. In: *Proc. 4th Internat. Symp. Pollin*, Maryland U.S.A., 49–57.
- Meyerhoff, G. (1954) Investigation on the effect of bee visits on rape. *Arch. Geflugelz Kleintierk* 3: 259–306.
- Mohr, N. A. and Jay, S. C. (1988) Nectar and pollen collecting behaviour of honeybees on Canola/Brassica campestris L. and Brassica napus L.). J. Apic. Res. 27: 131–136.
- Mosquida, J. J., Renard, G., Pellen-Delourme Pelleties, J. and Morice, (1987) Influence dessecretions nectarifers des lignes male streriles pour la production de. In: *Semences hybrids f. decalza*, INRA. France, 269–280.
- Ohsawa, R. and Nawal, H. (1988) Cross poolination efficiency of insect pollinators. *Japanese J. Breeding* **38**: 91–102.
- Panda, P., Rath, L. K., Padhi, J. and Panigrahi, D. (1995) Relative abundance and foraging behaviour of common bee species on niger in Phulbani District, Orissa, India. *Indian Bee J* 57: 10–14.
- Rana, R. S. (1989) Aggressive and hoarding behaviour of *Apis mellifera* and *Apis cerana* F. and their role in pollination of plum and apple bloom *Ph.D. Thesis*, Himachal Pradesh University, Shimla, India.
- Rao, G. M. (1991) Studies on the floral biology and pollination requirements of scented methi (*Trigonella corniculata* Linn.). *Indian Bee J.* **53**: 39–43.
- Rao, G. M. and Suryanarayana, (1990) Studies on the foraging behaviour of honeybees and its effect on the seed yield in niger. *Indian Bee J.* 52: 31–33.
- Renard, M. and Mesquida, J. (1987) Preliminary study on entomophilous pollination of male steriles rape in a mixed cultivation system comprising variable perportions of pollienizer plants. *Bul. Tech. Apicult.* 14: 99–104.
- Verma, L. R. and Dulta, D. K. (1986) Foraging behaviour of Apis cerana indica and Apis mellifera in pollinating apple flowers. J. Apic. Res. 25: 197–201.

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# Time of adult emergence and sex ratio of rice blue beetle, *Leptispa pygamaea* Baly (Coleoptera: Chrysomelidae) and extent of damage caused by the pest to two rice varieties

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ABSTRACT: The adult emergence and sex ratio of rice blue beetle. Leptispa pygmaea Baly was studied at Pattambi. Kerala during Kharif 2004–Rabi 2005. Highest emergence occurred between 8.00 and 11.00 h (25 and 15 per cent) and between 16.00 to 18.00 h (20 and 10 per cent). The female to male ratio during the peak period, June to November, in the field was 2.6:1. But during the lean period, the male beetles were more dominant in the field. Though both grubs and adults caused damage to rice; highest damage was inflicted by the grub followed by the adult female and male beetles irrespective of rice varieties. A grub consumed 82 per cent more leaf area than a male beetle while a female beetle consumed 33 per cent more leaf area than a male. The rice variety Jyothi was more susceptible than Aiswarya to the attack of Lpygmaea. © 2008 Association for Advancement of Entomology

KEYWORDS: rice blue beetle, *Leptispa pygmaea*, time of adult emergence, sex ratio, damage intensity

### INTRODUCTION

Severe outbreaks of rice blue beetle, *Leptispa pygmaea* Baly have been reported recently in the major rice growing tracts of northern districts in Kerala. The pest inflicted extensive damage during the early stages of rice in both *Kharif* and *Rabi* seasons. Since the blue beetle has so far been considered as a minor pest, no study on this pest has been carried out in Kerala. A study on the time of emergence and sex ratio of the pest and the nature and extent of damage caused would be helpful in deciding control strategies. Hence, the present study on these aspects was undertaken.

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### MATERIALS AND METHODS

The experiment was carried out in the net house under prevailing temperature (maximum  $30.1 \pm 1.4\,^{\circ}\text{C}$ ; minimum  $23.1 \pm 0.69\,^{\circ}\text{C}$ ) and relative humidity (94.33  $\pm$  2.11%) at the Regional Agricultural Research Station, Pattambi, Kerala Agricultural University during 2004–2005. For assessing the time of emergence of *L. pygmaea*, 15 days old potted rice seedlings (variety Jyothi) with 30 freshly pupated pupae of rice blue beetle each were collected from the field and were covered with polyester cage and kept in the net house. Three replications were maintained for each treatment with a total of 90 pupae. The emergence of adult beetle was observed at one hour interval between 0800 to 1800 h for a period of 3–4 days. The mean per cent emergence of adult beetles was worked out separately for each replication. For studying the sex ratio, male and female beetles were collected randomly by 10 net sweeps from the rice fields at monthly interval during the peak period of beetle activity from June to November, 2005.

To assess the extent of damage caused by L. pygmaea on different varieties of rice, single adult (male/ female) and grub was released separately on 15 day old potted rice seedlings of Jyothi (short duration) and Aiswarya (medium duration) covered with polyester cages (49 cm  $\times$  18 cm). Four replications were maintained for each stage of the beetle. Number of damaged leaves per hill was recorded at 9 days after release. The leaf area consumed by the adult (male, female) beetle and grub was assessed by graphical method.

### RESULTS AND DISCUSSION

The hourly emergence of L.pygmaea from 8.00 h to 18.00 h was 25.6, 20.0, 15.6, 5.6, 2.2, 0.0, 0.0, 7.8, 17.8 and 5.6 per cent (Table 1). Highest emergence of adult beetles (25.6 and 15.6 per cent) occurred between 08.00 and 11.00 h and thereafter, it was gradually reduced and reached 2.2 per cent between 12.00 and 13.00 h. There was no emergence during 13.00 to 15.00 h. The adults started emerging again from 15.00 h (7.8 per cent) and rose to 17.8 per cent between 16.00 and 17.00 h and ended with 5.6 per cent between 17.00 and 18.00 h. It is thus evident that the emergence of rice blue beetle was highest during morning and evening and no emergence occurred during the noon time when the temperature was at its peak indicating the influence of temperature on the adult emergence of rice blue beetle. The time of adult emergence has got much significance in chemical management of the pest as it would help in deciding the time of insecticide application preferably with the peak emergence of the beetle either in the morning or evening hours. No earlier work has been reported on the time of emergence of L. pygmaea. The present finding is in conformity with Swamiappan et al. (1990) who observed that the active mating of L. pygmaea occurred during morning and evening hours coinciding with the peak emergence of the adult beetles.

Adult beetle population recorded by monthly sweep net collections from the field during the peak period of activity from June'05 to November'05 were 185, 220, 211,

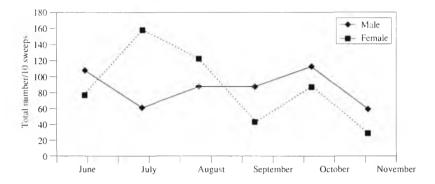


FIGURE 1. Average number of male and female beetles in the field (year 2005).

TABLE 1. Time of emergence of *L. pygmaea* during different hours of the day

Time	Number of	of adult beetles emerged
(Hours)	Mean	Per cent emerged in each interval
8.00-9.00	7.7	25.6
9.00-10.00	6	20.0
10.00-11.00	4.7	15.6
11.00-12.00	1.7	5.6
12.00-13.00	0.7	2.2
13.00-14.00	0.0	0.0
14.00-15.00	0.0	0.0
15.00-16.00	2.3	7.8
16.00-17.00	5.3	17.8
17.00-18.00	1.7	5.6

132, 202 and 90. The number of male/female population in the above catches were 108/77, 61/159, 88/123, 88/44, 114/88 and 60/30 (figure 1). Females predominated the field population during July and August when the total population was high and males were dominant during June, September, October and November, 2005 when the total field population was comparatively lower. This is in close agreement with the results of Dalvi *et al.* (1985).

Both the grub and adult of *L. pygmaea* feed on the upper surface of rice leaves by scraping chlorophyll matter, leading to longitudinal white streaks on them. The streaks made by the grub were found to be shorter and narrower as earlier reported by Patel and Shah (1985). The damage of blue beetle resembled the attack of rice leaffolder, *Cnaphalocrocis medinalis* except for the absence of webbing of leaves. In case of severe damage by *L. pygmaea* the rice leaves were seen folded longitudinally and ultimately dried up. From a distance, the damaged rice patch showed severe drying. Swamiappan *et al.* (1990) also observed that in certain pockets, when the young rice

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Rice variety	Stage of the beetle	Leaf damage (%)	Leaf area consumption (mm <sup>2</sup> )
Jyothi	Grub Adult female	67.09 28.27	3.82 1.04
	Adult male	13.65	0.70
Aiswarya	Grub	57.55	2.58
	Adult female Ault male	25.17 10.47	0.60 0.34

TABLE 2. Feeding intensity of *L. pygmaea* on rice varieties

plants were attacked by rice blue beetle, it resulted in stunting and severe drying symptoms and the incidence was higher in shaded areas.

Among the different stages of the rice blue beetle, grub caused the highest damage followed by adult female in both the varieties of Jyothi and Aiswarya (Table 2). Male beetles caused lowest damage to rice leaves. Grubs caused 67.09 and 57.55 per cent while male beetles caused 13.65 and 10.47 per cent leaf damage in Jyothi and Aiswarya respectively. The grub stage was found to cause 16.6 per cent increase in damage in Jyothi over Aiswarya thus indicating a higher susceptibility of Jyothi to *L. pygmaea*. In Jyothi, grub caused 80 per cent more damage than male beetle while female beetle caused 52 per cent more damage than the male. But in Aiswarya, grub feeding was 82 per cent more than that of the adult male beetle whereas the female beetle caused 58 per cent more damage than the male.

The same trend was observed in leaf area consumption also by the different stages of L. pygmaea. Leaf area consumption was more in Jyothi than in Aiswarya (Table 2). The grub consumed highest leaf area of 3.82 mm<sup>2</sup> in Jyothi and 2.58 mm<sup>2</sup> in Aiswarya. Adult female beetle consumed leaf area of 0.6 mm<sup>2</sup> in Aiswarya and 1.04 mm<sup>2</sup> in Jyothi .The consumption of male beetle was 0.34 mm<sup>2</sup> and 0.70 mm<sup>2</sup> in Aiswarya and Jyothi respectively. The cumulative consumption due to grub, male and female beetle was 5.56 mm<sup>2</sup> leaf area in Jyothi and 3.52 mm<sup>2</sup> in Aiswarya again confirming the higher susceptibility of Jyothi to the pest. It is thus proved that grub of L. pygmaea caused the maximum damage followed by female and male adult beetles irrespective of varieties in rice. A grub consumed 82 and 87 per cent more in Jyothi and Aiswarya respectively than that consumed by a male beetle, while the female beetle consumed 32.6 and 43 per cent more leaf area than that by a male in Jyothi and Aiswarva. In rice hispa, similar observations were reported by Deka and Hazarika (1997) who indicated that significantly more leaf area was consumed by females than males. Budharaja et al. (1979) observed that a single adult beetle of rice hispa consumed 25.3 mm<sup>2</sup> leaf area.

### REFERENCES

- Budharaja, K., Rawat, R. R. and Singh, O. P. (1979) Field efficacy and economics of some insecticides against *Dicladispa armigera* on paddy. *Indian Journal of Plant Protection* 7(2): 119–123.
- Dalvi, C. S., Dumbre, R. B. and Khanvilakr, V. C. (1985) Bionomics, seasonal incidence and off season biology of rice blue beetle. *Journal of Maharashtra Agricultural Universities* 10(2): 185–187
- Deka, M. and Hazarika, L. K. (1997) Mating and oviposition behaviour as affected by differences in sex ratio of rice hispa. *Crop Research Hisar* **13(1)**: 163–167.
- Patel, C. B. and Shah, A. H. (1985) Biology of rice blue beetle, *Leptispa pygmaea* Baly. (Coleoptera: Hispinae). *Bulletin of Entomology* **26(2)**: 120–128.
- Swamiappan, M., Rajaram, K. S. and Nair, V. C. (1990) Outbreak of blue beetle in India. IRRI News letter 15(2): 16–17.

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## Histomorphological derangements in the ovary of *Oryctes rhinoceros* (Coleoptera: Scarabaeidae) treated with methanolic extract of *Annona squamosa* leaves

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ABSTRACT: Topical application of methanolic extract of *Annona squamosa* leaves caused histomorphological aberrations in the ovary of the adult *Oryctes rhinoceros* affecting full growth of ovum. © 2008 Association for Advancement of Entomology

KEYWORDS: Oryctes rhinoceros, Annona squamosa, effect of plant extract, ovary

### INTRODUCTION

Phytochemicals are known to inhibit reproduction in various Orders of insects. Shalom et al. (1988) found abnormal oocyte development in Locusta migratoria by azadirachtin. Acorus calamus extracts resulted in decreased reproductive potential in Tribolium castaneum (Joseph et al., 1994). Essential oils from the rhizomes of Acorus calamus were toxic to the eggs and induced significant reduction of oviposition in Callosobruchus phaseoli (Rahman and Schmidt, 1999). Limonoids from neem, Azadirachta indica and methanol extracts of leaves from the Indian white cedar Dysoxylum malabaricum showed anti ovipositional activity in Anopheles stephensi (Nathan et al., 2005, 2006). The present study was undertaken to assess the effect of methanolic extract of the leaves of Annona squamosa (Annonaceae) on the ovary of the coconut rhinoceros beetle, Oryctes rhinoceros.

### MATERIALS AND METHODS

Adults of *Oryctes rhinoceros* were obtained from late 3rd instar larvae collected from local dung pits and reared in the laboratory in sterilized cow dung. Adult eclosion date was noted to determine the age of the beetles. One day old beetles were used for the treatment. Experiment and control groups consisted of seven beetles each.

The beetles were kept individually in glass containers with wide mouth. They were fed on ripe banana pieces and checked twice a day to monitor survival. Mature leaves of A. squamosa were dried under shade, powdered and extracted successively using methanol. After evaporation of the solvent, 0.1% stock solution of the solute was prepared, by dissolving 100 mg of it in 100 ml methanol. Using a Hamiltons microlitre syringe 10  $\mu$ l of the stock solution (10  $\mu$ g Annona extract) was topically applied, on the ventral side of the abdomen to see whether there is any direct effect on the ovaries. Controls received an equal volume of methanol. Treated and control insects were dissected in cold insect Ringer under binocular dissection microscope, on the 34th day as it was understood from preliminary studies that vitellogenesis and choriogenesis were completed at least in the proximal oocytes of all ovarioles by 32-34 days. Morphological observations were made in unfixed tissues under dissection microscope. Ovarioles were fixed in aqueous Bouins fixative for 48 h. Following standard histological procedures, paraffin sections, 5  $\mu$ m thick, were stained in Ehrlich's Haematoxylin and Eosin, mounted in DPX and examined under the light microscope.

### RESULTS AND DISCUSSION

Morphological variations were observed in the ovarioles of the treated beetles. In treated beetles, the terminal filaments are very much reduced or absent, and as a result the germaria floated free in the body cavity. In the control, terminal filaments are long and interconnected at their tips. Germarium in the treatment is short; not intact and is easily rupturing. The vitellarium is reduced, formed of 2 or 3 fragile follicles, while in the control there are 5–7 follicles in various stages of maturation; the proximal one being the most developed. In the treated beetles, the proximal and penultimate follicles show abnormal shape and look slightly compressed. Occasionally, the proximal and penultimate follicles appear fused. Anterior to the penultimate follicle, one more follicle is seen in some ovarioles. Follicles did not show inner, dark granules seen in the control. The interfollicular tissue which is distinct in the control, is indistinct in the treated beetles (figure 1a, b).

Ovarioles of the treated beetles also showed histological abnormalities. Germarium of control is almost uniformly quiescent and passive composed of trophocytes with indisinct cell boundaries. In the treated beetles, the germarium is degenerated; a major part of it is diffused and vacuolated. There are a few trophic nuclei in the diffused mass. Basal part of the germarium consists of prefollicular cells and a few pro oocytes in the controls, while in the treated lot, the prefollicular region is empty, vacuolated and distorted.

Vitellarium is highly abnormal in treated beetles, consisting of 3–4 abnormal shaped follicles looking as if some of them are fused. The interfollicular tissue is not differentiated. In all follicles, follicular epithelium is thin, disrupted and not properly oriented; follicle cells are deformed; vitellogenesis and choriogenesis are arrested (figure 2). Vitellarium of the control consists of 5–7 follicles of which the anterior one or two are in pre/early vitellogenesis and the remaining ones in progressive stages of

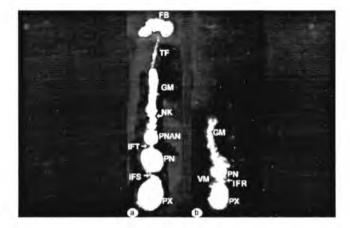


FIGURE 1. Morphology of ovariole (a) control (b) treated.

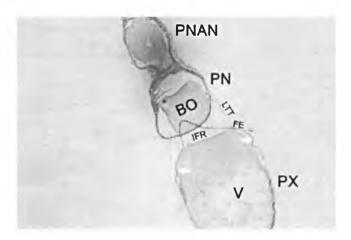


FIGURE 2. L.S. of the vitellarium of the treated beetle ( $\times 100$ ).

vitellogenesis. Each follicle is formed of an oocyte and a layer of follicular epithelium surrounding it. Follicular epithelial cells of the vitellogenic follicles are big, round and are actively engaged in the transfer of vitellogenic materials into the oocyte. The interfollicular tissue is formed of thin walled cells of varying shapes, generally round or spindle shaped. It is continuous with the follicular epithelium of the succeeding and preceding oocytes. On either side of the inter follicular tissue the inner layer of the ovariole sheath is expanded and modified into a trophic tissue, named in this study as the lateral trophic tissue (Fig.3). In the proximal oocyte of the control, vitellogenesis is completed and yolk is uniformly distributed in the form of globules of varying size and as fine granules. After the completion of vitellogenesis a chorion is deposited by the follicle cells. The proximal oocyte of treated beetles is almost flattened with irregular

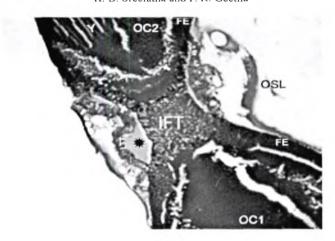


FIGURE 3. L.S. of vitellogenic follicle of control beetle (×200).

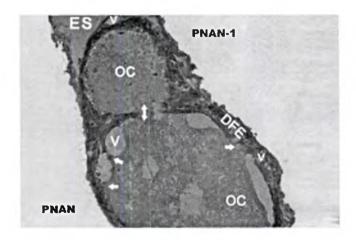


FIGURE 4. L.S. of vitellogenic follicle of treated beetle (×200).

TF. Terminal filament; GM, Germarium; VM, Vitellarium; IFS, Interfollicular stalk; IFT, Interfollicular tissue; IFR, Inter follicular region; FE, Follicular epithelium; PX, Proximal oocyte; PN, Penultimate; PNAN, Anterior penultimate; Y, Yolk; LTT, Lateral trophic; BO, Block of; DFE, Defective follicular epithelium; V, Vacuoles; FB, Fat body; ES, Empty space; OC1 & 2, Vitellogenic ic oocytes; OSL, Outer layer of ovariole sheath.

Arrows indicate transfer of vitellogenic materials from LTT, IFT and FE towards the oocytes in figure 3, and bulging of ooplasm in figures 2 and 4.

Double headed arrow indicates continuity of the ooplasm of the oocytes, IFT being absent. Asterisk indicates secretions accumulated between LTT, IFT and FE.

borders and cytoplasmic protrusions. Ooplasm looks reticulate due to vacuolation and shrinkage. The follicular epithelium is thin and a major part of it is lifted off from the oocyte surface. The penultimate oocyte is compressed; ooplasm appears

as a single block and empty spaces are seen between the oocyte and the follicular epithelium. Cellular integrity of the follicular epithelium is not evident, but the layer holds some secretion which is in contact with the ooplasm. There is no interfollicular tissue between proximal and penultimate oocytes, the region being empty. The lateral trophic tissue is thin and membraneous (figure 2). Ooplasm of the follicles which are situated anterior to the penultimate one shows vacuolation and bulging towards the follicular epithelium. These oocytes are continuous since there is no inter follicular tissue in between. Empty spaces are seen in front of the follicles and also between the follicular epithelium and the oocyte (figure 4).

Morphological variations observed in the present study due to Annona leaf extract treatment includes: reduced size of ovarioles, reduced number of follicles and absence of inter follicular tissue. Ovariole size, length and oocyte number were reduced in *Dysdercus cingulatus* treated with *Eupatorium odoratum* and *Vitex negundo* extracts (Prameela, 1997). Reduced ovary size and oocyte growth were observed in *Ceratitis capitata* treated with *Annona squamosa* seed extract (Epino and Chang, 1993) and in neem fed females of *Epilachna varivestis* (Schulz, 1981). Ghazawi *et al.* (2007) reported shrinkage of ovaries, oocyte growth inhibition and reduced number of eggs in the grasshopper *Heteracris littoralis* treated topically with azadirachtin. Methanolic neem seed kernel extract inhibits ovary development and oocyte differentiation in *Trogoderma granarium* (Chellayan and Karnavar, 1990a). In the treated ovary of *O. rhinoceros* the terminal filaments are either very short and free or absent. Free and short terminal ends of ovarioles and reduced ovariole length were reported in *Corcyra cephalonica* emerged from neem fed larvae (Chanda and Chakravorty, 2000).

The extract induced high degenerative changes in the germarium involving vacuolation in the trophic and pre follicular regions. Presence of empty spaces and vacuolated cytoplasm might be due to the cellular degeneration leading to the reduction in the number of cells. Prameela (1997) reported degeneration of trophocytes in the ovary of *D. cingulatus* treated with extracts of *Chromolaena odorata* (syn. *Eupatorium odoratum*) and *Vitex negundo*. Abnormal shape and size of the follicles is apparently due to the defective orientation of the follicular cells. The follicular cells fail to surround the oocyte as an intact, continuous layer. Large vacuoles amidst the ooplasm might have resulted from ooplasmic shrinkage.

In the present study the follicle cells are abnormal and deformed. It seems that the follicle cells are not competent to take over their normal functions during vitellogenesis. Yolk is absent in the follicles. Impairment in the follicle cell differentiation and functioning remains a significant reason for arrested vitellogenesis observed in the treated insects. In the control, the interfollicular tissue and lateral trophic tissue also have active roles in the transfer of vitellogenic materials into the oocyte. Follicular epithelium, inter follicular tissue and lateral trophic tissue are atrophied or absent in all treated insects. These structural abnormalities have a cumulative role in the present results associated with arrested vitellogenesis and oocyte development in *O. rhinoceros*. Disrupted and irregular follicular epithelium, abnormal vitellogenesis and vacuolated ooplasm were observed in *Heteracris littoralis* by azadirachtin (Ghazawi *et al.*, 2007).

Thin and abnormal follicular epithelium, irregular shape of oocytes, arrested vitellogenesis and deformed ovarioles were reported in *Dysdercus cingulatus* by methanolic extract of *Chromolaena odorata* (Prameela, 1997). Structural and functional anomalies induced by the phytochemicals in the follicular epithelial cells are possibly the reason for the failure of chorion deposition.

### ACKNOWLEDGEMENT

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### REFERENCES

- Chanda, S and Chakravorty, S. (2000) Effect of neem mixed larval food on development of reproductive organs and nature of fertility of the emerged morphs of *Corcyra cephalonica* (Stainton) (Lepidoptera: Pyralidae). *Ann. Entomol.* 18: 19–26.
- Chellayan, S. and Karnavar, G. K. (1990a) Influence of neem kernel extract on morphogenesis and vitellogenic oocyte development in *Trogoderma granarium*. Everts. Proc. Indian. Acad. Sci. (Anim Sci) 99: 113–118.
- Epino, P. B. and Chang, F. (1993) Biological effects of *Annona squamosa* L. seed extracts against the Mediterranean fruitfly *Ceratitis capitata* (Wiedmann) (Diptera: Tephritidae). *Philippine Entomol* 2: 214–217.
- Ghazawi, N. A., El-Shranoubi, E. D., El-Shazly, M. M. and Abdel Rahman, K. M. (2007) Effects of azadirachtin on mortality rate and reproductive system of the grasshopper *Heteracris littoralis* ramb. (Orthoptera: Acrididae). *Journal of Orthoptera Research* 16: 57–65
- Joseph, M., Mukherjee, S. N. and Sharma, R. N. (1994) Growth inhibition and impairment of reproductive potential in *Tribolium castaneum* (Herbst.) (Coleoptera: Tenebrionidae) by commercially available plant extracts. *Insect Sci. Appl.* 15: 197–202.
- Nathan, S. S., Kalaivani, K. and Murugan, K (2005) Effects of neem limonoids on malaria vector *Anopheles stephansi* Liston. (Diptera: Culicidae). *Acta Tropica* **96(1)**: 47–55.
- Nathan, S. S., Kalaivani, K. and Kim, Sehoon (2006) Effects of *Dysoxylum malabaricum* Bedd. (Meliaceae) extract on the malarial vector *Anopheles stephansi* Liston. (Diptera: Culicidae). *Bioresource Technology* **97**: 2077–2083.
- Prameela, M. (1997) Bioinsecticides in the physiology of red cotton bug *Dysdercus cingulatus* Fabr. (Heteroptera: Pyrrhocoridae). *Ph. D Thesis*, University of Kerala.
- Rahman, M. M. and Schmidt, G. H. (1999) Effect of *Acorus calamus* (L.) (Araceae) essential oil vapours from various origins on *Callosobruchus phaseoli* (Gyllenhal) (Coleoptera: Bruchidae). *Journal of Stored Products Research* 35: 285–295.
- Schulz, W. D. (1981) Pathological alterations in the ovaries of *Epilachna varivestis* induced by an extract from neem kernels. In: *Proc. 1st Int. neem Conf. (Rottach Egern)* 1980, 81–96.
- Shalom, U., Applebaum, S. W. and Pener, M. P. (1988) Vitellogenesis and oocyte development in azadirachtin induced fifth-instar overage nymphs of *Locusta migratoria* (L.). *Arch. Ins. Biochem. Physiol.* **9**: 313–322.

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Article No. ent.33204



### Distribution pattern of white fly, *Bemisia tabaci* under natural condition on okra cultivars

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ABSTRACT: Spatial distribution of *Bemisia tabaci* studied on okra cultivars showed significant differences on the density of pest at different stages of crop growth i.e. 30, 45 and 60 days after planting. The peak population was seen on 60 day old crop in 2005 and 2006, while the lowest was on 30 day old crop. Population density was higher (3.2–6.7 adult/leaf) during kharif season of 2006 than that of 2005 count (2.1–4.2 adult/leaf). *B. tabaci* followed a regular distribution while aggregated distribution pattern was also recorded when the population was low in 2005. © 2008 Association for Advancement of Entomology

KEYWORDS: Spatial distribution, Abelmoschus esculentus, Bemisia tabaci

### INTRODUCTION

White fly, *Bemisia tabaci* (Gennadius) [Homoptera: Aleyrodidae] is polyphagous pest feeding on an estimated 6000 plant species and causes enormous direct damage to crops and also serves as vectors of over 100 plant viruses (Costa, 1976). It is responsible for transmission of yellow mosaic virus (YMV), which is a major constraint for cultivation of the okra crop in India (Neerja et al., 2004). The distribution of insects in their habitat follows characteristic patterns depending on their inherent behaviour (Iwao, 1970) and this behaviour will affect population density estimates which constitute an essential component in insect management programmes. Present investigation was hence undertaken to understand the distribution pattern of *B. tabaci* on okra varieties.

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TABLE 1. Spatial distribution of B. tabaci on okra (year 2005).

Variety	Mean density per leaf Variance (52)	sity per	leaf	Varian	ce (52		riance m	can ratio (S <sup>2</sup> /m	() Lloyd	's Index o.	Variance mean ratio (\$2^m)   Lloyd's Index of Mean Crowding   Lloyd's Patchiness Index (X)   Dispersion Parameter (X)   David and Moore's Index	Lloyd's	Patchines	s Index (X)	Dispers	ion Paran	neter (K)	David and	Moore's	Index
	30 4	5	60 3	30 4	45 6		30 45	09	30	45	09	30	45	09	30	45	09	30	45	09
	DOC DOC		DOC DO	DOC DO	DOC DO	DOC DO	DOC DOC	DOC 3	DOC	DOC	DOC	DOC 1	DOC	DOC	DOC	DOC	DOC	DOC	DOC	DOC
Arka Anamica 2,768 2,734	2.768 2.7		3,430 1.0	1.077 1.9	1.987 2.3	2.254 0.3	0.389 0.726	6 0.657	3.080	3.320	4.015	0.779	0.726	0.900	4.531	10.00	10,000	119:0-	-0.274	-0.343
Parbhani	2,180 2,244		2,248 0,6	0,613 0,9	0.960	3,506 0,2	0,281 0.428	655.1 8	2.400	2.586	2.516	0.671	0.744	0.751	3.033	3.921	4.017	617.0-	-0.572	-0.559
Karanti																				
Selection-9	2.892 2.152		2.637 0.9	0.955 0.4	0.416 0.7	0.399 0.3	0.330 0.193	3 2.420	3.130	2.306	2.757	0.768 0	0.625	0.688	4.317	2,667	3.105	029'0-	-0.807	-1.420
Seed Tech	2,126 2,118		3,278 0.4	0.465 0.1	0.157 1.3	1.338 0.2	0.218 0.074	4 0.408	2.300	2.178	3,605	0.633 (	0.563	0.820	2,721	2.287	5,538	-0.782	-0.926	-0.592
-71																				
NBR-1	2.184 2.120		2.508 0.2	0.287 0.3	0.337 2.0	2.006 0.1	0.131 0.159	664.0	2 290	2.247	3.148	0.603 0	0.604	0.921	2.514	2.520	12.529	-0.869	-0.841 -0.201	-0.201
Ajcet	3.740 3.508		4,016 0.5	0.570 3.0	3.030 5.0	5.098 0.1	0.152 0.864	4 7.269	3.860	4.198	5.032	0.774	0.942	0.933	4.422	25.63	14,905	0.848	-0.136	-6.269
-311 (F1)																				
Hari Rani	3,292 2.71	2	2.720 1.0	1.095 0.2	0.286 2.3	2.324 0.3	0.333 0.105	5 0.854	1.700	2.796	3,403	0.797 0	1/9'0	0.907	4.926	3.032	10.784	1990-	-0.895	-0.854
Depti	2.672 3.318		2.188 0.3	0.383 0.4	0.418 1.2	1.295 0.1	0.143 0.126	6 0.592	2.780	3.419	2.662	0.680	0.737	0.814	3.118	3.796	5,360	-0.857	-0.874	-0.592
Green Gold	3.134 3.046		4,228 0.4	0.417 2.0	2.093 2.2	2,220 0.1	0.133 0.687	7 0.525	3,240	5.365	4.648	0.724 0	0.897	0,888	3.615	9.685	8.906	198.0	-0.313	-0.525
Aject-121	2.234 2.472		3.558 0.3	0.303 0.3	0.345 0.8	0.854 0.1	0.136 0.139	9 0.240	2.340	2.583	3.750	0.613 0	0.652	762.0	2,584	2,873	4.683	-0.864	198.0-	-0.240

DOC = Day Old Crop

When variance mean ratio is less than 1, 1 or more than 1, distribution will be regular, poisson or aggregated respectively.
 If patchiness value is less than 1, equal, or more than 1, the distribution will be dispersed, random or clumped, respectively.

3. Higher value of exponent *K* indicates the greater extent of aggregation and it is a negative binomial.

4. When David and Moore's Index value is zero (0), the distribution is random, positive value is for negative binomial (aggregated) and negative for positive binomial (regular).

TABLE 2. Spatial distribution of B. tabaci on okra (year 2006)

Variety	Mean	density	Mean density per leaf		Variance (S2)		Varian	ce mean	Variance mean ratio (\$2/m). Lloyd's Index of Mean. Lloyd's parchiness. Dispersion parameter (K). David and Moure's index	Lloyd	s Index	of Mean	Lloyd	's patch	ness	Dispersi	оп рагап	ieter (K)	David m	Moone	s index
											crowding	500	E	index (X)							
	30	45	09	30	45	09	30	45	09	30	45	09	30	54	09	30	45	09	30	45	()9
	200	1000	DOC	DOC	DOC	D00	DOC	DOC	DOC	DOC	DOC	DOC	DOC	D0C	DOC	DOC	DOC	DOC	DOC	DOC	DOC
Arka Anamica	3.724	3.906	4.974	0.977	2.247	3.193	0.262	0.572	0.642	3.934	4 366	5.487	0.981	0.892	0.928	5.048	9.196	13,891	0.738	-0.428	-0,358
Parbhani Karanti	5.37N	5.198	5.412	0.432	4,393	5.627	0.080	0.845	1.039	6.085	5.870	6.240	0.829	176.0	0.993	5.847	33.56	13.820	0.920	-0.155	-0.039
Selection-9	4.650	4,000	3.989	2,890	7,667	3,095	0.622	1.917	0.776	3.087	5 533	4.605	6160	0.771 0.934	0.934	12 284	4.363	17.850	-0.388	-0.917	-0.226
Seed Tech - 71	3.196	3.858	6.520	1,998		5.313 0.352	0.625	1.377	0.054	4 600	4.959	9.560	0.999	0.903 0.855	0.855	8.526	10.40	6.863	-0.375	-0.377	-0.946
NBR - I	3.526	5.182	5.148	1.783		1,800 2,667	0.506	0.347	0.518	3,390	5.459	5.560	0.879	0.874 0.906	906.0	8.280	7.939	10.681	0.494	-0.653	-0.482
Ajeer 311 (F1)	5.130	6.438	6.716	3.525	4,560	0.067	0.687	0.708	0.108	\$.679	7,004	6.790	0.939	0.955	998.0	16.397	22.06	7.459	-0.313	-0.292	0.892
Hari Rani	3.324	4,174	5.356	1.535	6.837	6.837 3.273	0.462	1.638	0.611	4.636	5.484	5 840	0.972	0.847	0.912	6.175	6.542	11.372	0.538	-0.638	-0.389
Depti	4.164	4 992	3.928	2.785		3,350 3,499	8990	0.671	0.891	7.259	5.528	4 630	0.921	0.934	6260	12.571	15.17	32.210	-0.332	-0.331	-0.109
Green Gold	3.324	5.180	4.384	1.730	0.703	5.787 0.520		0.136	1.320	3.740	5.288	5 440	0.856	0.833	0.927	6.932	5.993	13,698	-0.480	-0.864	-0.320
Ajeet-121	5.658	5.574	₹ 764		0.947 1.145 1.375 0.167 0.205	1.375	0.167	0.205	0,365	5,792	5.738	4 050	0.853 0.857 0.832	0.857	0.832	6.796	7,015	5.930	-0.833 -0.795 -0.635	-0.795	-0.635

DOC = Day Old Crop

When variance mean ratio is less than 1, 1 or more than 1, distribution will be regular, poisson or aggregated respectively.

2. If patchiness value is less than 1, 1 or more than 1, distribution will be dispersed, random or clumped, respectively.

 Higher value of exponent K indicates the greater extent of aggregation and it is a negative binomial.
 David and Moore's Index value for zero (0) a random, the distribution is random, positive value is for negative binomial (aggregated) and negative for positive binomial (regular).

### MATERIALS AND METHODS

The experiments were carried out in the field of Faculty of Agricultural Sciences, Aligarh Muslim University, Aligarh, during kharif season of 2005 and 2006. Seeds of okra cultivars (vide Table 1) were sown in 3rd weak of Feb. 2005 and 2006 in  $3 \times 3$  m plots with a row spacing of 45 cm and plant to plant spacing of 30 cm. Each variety was replicated in three plots. Standard agronomic practices were followed. Pest population on 30, 45 and 60 days old crops (DOC) were assessed. Five plants were selected at random from each plot and tagged. Counting was done in morning hours and a total of six leaves (2 leaves each from top, middle and lower position) were used for the same. The data were analyzed statistically. The spatial distribution pattern was assessed by calculating variance mean ratio (Elliot, 1977), Lloyd's index of Patchiness (Lloyd's, 1967), Lloyd's index of mean crowding (X), index of clumping of David and Moore (1954) and Dispersion parameter (k) (Katti and Gurland, 1962).

### RESULTS AND DISCUSSION

The density of *B. tabaci* population on the leaves of okra plant exhibited a significant difference on crops at three different growth stages viz., 30, 45 and 60 day old crops (Table 1, 2). The peak population was observed on 60 day old crop in both years and lowest on 30 day old crop. The population density was higher (3.2–6.7 adult/leaf) during 2006 as compared to 2005 (2.1–4.2 adult/leaf). The highest population (6.72) was recorded on variety Ajeet–311 at 60 day old crop in kharif season of 2005. The variance was less than mean at all observations except some cases on 60 day old crop of 2005 (Parbhani Karanti, Ajeet-311) and 45 day (Selection-9, Hari Rani) and 60 day old crop of 2006 (Green gold), which indicated a regular distribution.

Variance mean ratio was less than 1 at all observations except some cases where regular distribution and aggregated nature was noticed on 45 day old crop during 2006 (Selection-9, Seed Tech-71, Hari Rani), 60 day (Green Gold) and 60 day old crop of 2005 (Parbhani Karanti, Selection-9 and Ajeet-121).

The patchiness index was not significantly greater than 1, which showed a regular distribution nature of *B. tabaci*. The distribution pattern of population of *B. tabaci* can be adequately explained by the exponent 'K' of the negative binomial, which is an index of aggregation of population. Southwood (1978) reported that the smaller the value of 'K', the greater the extent of aggregation. The value of 'K' in all observations was more than two (2) and this indicates that the extent of aggregation was less and the distribution approach randomness or regular dispersion. The index of clumping of David and Moore gave a value of zero for a random population and the values with negative and positive signs showed a regular and contagious pattern of insect population respectively. The observed values of David and Moore's index were lower than the random values and are having negative signs which again shows regular nature of the pest in most of observations. The value of mean crowding increased with the increase in mean adult density. Values of Lloyd's index of Patchiness were always less than 1 in the present study which further confirms the regular nature of distribution of

white fly. Sharma (2001) observed that the pest followed the regular distribution in the beginning of the infestation each year and contagious distribution during rest of the period of crop infestation. Pereira et al. (2004) found a positive binomial distribution of adult of B. tabaci biotype B in common bean, Phaseolus vulagaris in November and February plantings. The results of Rathore and Tiwari (1998) showed an aggregated distribution of B. tabaci on mung bean (Vigna radiata (L.) Wilczek), urd bean (Vigna mungo (L.) Hepper) and cowpea (Vigna unguiculata (L.) Walp.) during the summer and kharif seasons. They also reported that crops, cropping stage and seasons did not affect the aggregated behaviour. However, the degree of aggregation was greater when the population was high and showed a tendency towards randomness in the case of low density of B. tabaci. Shen et al. (2005) found that the number of B. tabaci adults was highest on the upper, tender and fully open leaves on aubergine, watermelon and musk melon and the pest was aggregated on these plants as well as on cucumber. Therefore, it can be concluded that the distribution pattern of adult population of B. tabaci followed regular pattern of distribution on okra crops though aggregated distribution pattern was recorded on many others.

### REFERENCES

- Costa, A. S. (1976) Whitefly-transmitted plant diseases. *Annual Review of Phytopathology* **14**: 429–449.
- David, F. N. and Moore, P. G. (1954) Notes on contagious distributions in plant populations. *Annals of Botany* 18: 47–53.
- Elliot, J. M. (1977) Some methods of statistical analysis of samples of Fenthic invertebrates. In: Freshwater Biological Association Scientific Publication No. 25, 121.
- Iwao, (1970) Analysis of spatial pattern of animal populations prograess of resesch in Japan. *Japanese Revive* 3: 41–54.
- Katti. S. K. and Gurland, J. (1962) Efficiency of certain methods of estimation for the negative binomial and the Neyman type A distribution. *Biometrika* 49: 215–226.
- Lloyd, M. (1967) Mean Crowding. Journal of Animal Ecology 36: 1-30.
- Neerja, G., Vijaya, M., Chiranjeevi, C. and Gautham, B. (2004) Screening of okra hybrids against pest and diseases. *Indian Journal of Plant Proection.* **32(1)**: 129–131.
- Pereira, F. A., Marcelo, J. R., Boica, L., Arlindo, C., Barbosa, and Jose, (2004) Spatial distribution of *Bemisia tabaci* (Genn.) biotype B (Hemiptera: Aleyrodidae) on bean crop (*Phaseolus vulgaris* L.). *Neotropical Enomology* **33(4)**: 493–498.
- Rathore, Y. S. and Tiwari, S. N. (1998) Influence of crops and cropping seasons on spatial distribution of *Bemisia tabaci* Genn. *Indian Journal of Pulses Research* 11(2): 76–85.
- Sharma, K. (2001) Spatial distribution of whitefly (Bemisia tabaci) on upland cotton (Gossypium hirsutum). Indian Journal of Agricultural Sciences 71(6): 424–426.
- Shen, B. B., Ren, S., Musa, P. D. and Zhou, J. (2005) The spatial distribution pattern of *Bemisia tabaci* adults. *Chinese Bulletin of Entomology* **42**(5): 544–546.
- Shukla, R. P. (1986) Spatial pattern of egg plant shoot and fruit borer, *Leucinodes orbonalis* Guen. (Lepidoptera: pyraustidae) larvae. *Insect Science and its Application* **7(5)**: 611–615.
- Southwood, T. R. E. (1978) *Ecological Methods*, English Language Book Society and Chapman and Hall, 524.



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## New reports of predatory mites (Acari: Prostigmata, Mesostigmata) from medicinal plants of Darjeeling district, West Bengal, India with description of a new species

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ABSTRACT: Eighteen species of mites under 6 families and 2 orders, recorded for the first time from 11 different species of medicinal plants of Darjeeling district of West Bengal, India, are reported in this paper. *Erythraeus cinchoni* sp. nov. is described and illustrated. © 2008 Association for Advancement of Entomology

KEYWORDS: Predatory mites, medicinal plants, new reports, *Erythraeus cinchoni* sp. nov.

### INTRODUCTION

Predatory mites are of great economic importance as they are used in the biological control of economically injurious mites (Slone and Croft, 2001). A systematic survey was conducted in different medicinal plant gardens, situated at different altitudinal ranges, extending from 594 ft (Sukna) to 6000 ft. (Darjeeling Town) of Darjeeling District, West Bengal during the months of November-December, 2006. Special attention was made to collect mite specimens from the Cinchona plantations at Mungpoo at an altitude of 3,500 ft. Altogether 20 species of mites were identified from 11 different species of medicinal plants, all of which were predatory in nature except the one belonging to the genus *Tetranychus*. The present communication includes new reports of 18 species of mites belonging to 5 families and 2 orders. Besides, 5 species namely *Anystis baccarum* (Linnaeus), *Cunaxa myabunderensis* Gupta & Ghosh. *Agistemus unguiparvus* Gonzalez-Rodriguez, *Amblyseius* (*Amblyseius*) aerialis (Muma)

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and *Typhlodromus* (Amblydromella) himalayensis Gupta were recorded for the first time from the state of West Bengal. This paper also deals with the description and illustration of a new species of the genus *Erythraeus* Latreille (Family: Erythraeidae). Type specimens are kept in the Entomology and Wildlife Biology Research Laboratory, University of Calcutta, which in due course will be deposited in the National collection of Zoological Survey of India, Kolkata. All the measurements are given in microns. The senior author did the entire collection.

### Order I. Prostigmata

### Family 1. Anystidae Oudemans, 1902

### 1. Anystis baccarum (Linnaeus)

1758. Acarus baccarum Linnaeus, Systema Naturae, 10th Ed. 106.

Collection records: 1 Female, India: West Bengal, Darjeeling, Mungpoo Govt. Quinine Factory Plantation area, ex *Cinchona officinalis* Linn. dated: 21.11.2006.

Habitat (Medicinal plant): Cinchona officinalis (new report).

Distribution: India: Jammu & Kashmir, Punjab (Gupta, 2002); West Bengal (new report); Elsewhere: U.S.A., Juan Fernandez Island. (Close to Chile), St. Helena, Faeroes Island., Mexico, Australia, Europe, Japan, North and South Africa (Gupta, 2002).

### Family 2. Cunaxidae Thor, 1902

### 2. Cunaxa myabunderensis Gupta & Ghosh

1980. Cunaxa myabunderensis Gupta & Ghosh, Rec. Zool. Surv. India, 77: 190-192.

Collection records: 3 females, India: West Bengal, Darjeeling, 5th mile-Siliguri, ex *Zingiber* sp., dated: 23.11.2006.

Habitat (Medicinal plants): Aegle marmelos (Gupta, 2002), Zingiber sp. (new report).

Distribution: India: Andaman & Nicobar Islands. (Gupta, 2002), West Bengal (new report).

### 3. Cunaxa womersleyi Baker & Hoffmann

1948. Cunaxa womersleyi Baker & Hoffmann, An. Esc. Nac. Cienc. Biol. Mexico, 6: 234–235.

Collection records: 2 females, India: West Bengal, Darjeeling, Sukna, ex *Ocimum sanctum* L., dated: 24.11.2006.

Habitat (Medicinal plant): Ocimum sanctum (new record).

Distribution: India: West Bengal; Elsewhere: U.S.A. (Gupta, 2002);

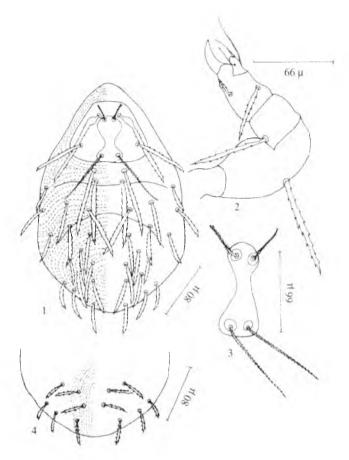


FIGURE 1. *Erythraeus cinchoni* sp. nov. (Larva): 1. Dorsal view; 2. Dorsal aspect of terminal segments of palp; 3. Crista metopica (enlarged view); 4. Venter of opisthosoma.

### Family 3. Erythraeidae Robineau-Desvoidy, 1828

### 4. Erythraeus cinchoni sp. nov. (figures 1-10)

### Larval diagnosis

Body oval, 495 long (from base of gnathosoma upto posterior tip of body) and 191 wide. Crista metopica 66 long and 33 wide, reaching anteriorly upto 1st pair of dorsal setae and little behind coxa II. Anterior sensillary short and 26 long, posterior sensillary 66 long. Pedipalp 165 long; tibial claw 26 long; seta on palp femur 66 long, seta on palp genu 50 long. Terminal seta on palp tarsus 43 long and the other two being 20 and 13 long respectively. Propodosomal scutum 92 long and 72 wide. Anterior dorsal propodosomal seta 116 long and posterior dorsal propodosomal seta 63 long- all being thick and serrate. Dorsal hypostomal setae 17 pairs long, thick serrate

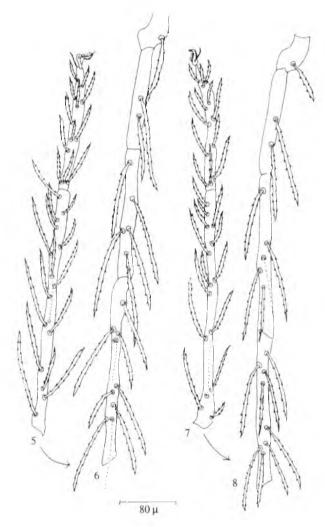


FIGURE 1. Erythraeus cinchoni sp. nov. (Larva): 5. Leg I- tarsus, tibia; 6. Leg I- genu, femur, Trochanter, coxa; 7. Leg II- tarsus, tibia; 8. Leg II- genu, femur, Trochanter, coxa.

and their length varies from 33–66. Ventral setae 12 pairs of which 2 pairs in mid ventral region about 20 long. Anteriorly, ventral setae of the propodosomal region less thick and less barbed. Postventral seta varies from 33–56 long. Hysterosomal striation transverse medially and oblique laterally; the posterior region striation mostly transverse. Setae of genitoanal region also long, serrate like those of dorsal region. Legs 3 pairs, being enormously long measuring Leg I 1006, Leg II 999 and Leg III 1013 long. Leg chaetotaxy as figured. All the leg setae being thick, serrate and long; tarsus terminates in a pair of claws.



FIGURE 1. Erythraeus cinchoni sp. nov. (Larva): 9. Leg III- tarsus, tibia; 10. Leg III- genu, femur, Trochanter, coxa.

Male: Unknown

Material examined

*Holotype*: Larva, India: West Bengal, Darjeeling, Mungpoo Govt. Quinine Factory plantation area, ex *Cinchona officinalis* Linn., dated: 21.11.2006, coll. Indranil Roy.

Paratypes: 3 Larvae, same data as for holotype.

Etymology: The species name is after the host plant.

### Remarks

This new species is close to *Erythraeus plumosus* (Khot, 1963) but differs from the structure of crista metopica and dorsal idiosomal setae which are extremely long in case of this new species. Further it can be differentiated from *E.* (*Zaracarus*) sibuljinicus (Haitlinger, 2004) by nature and number of setae on dorsal surface and also by shape of crista metopica.

### Family 4. Stigmaeidae Oudemans, 1931

### 5. Agistemus simplex Gonzalez-Rodriguez

1965. Agistemus simplex Gonzalez-Rodriguez, Univ. Calif. Pub. Ent., 33-34.

Collection records: 2 females, India: West Bengal, Darjeeling, Salugara, ex *Zingiber* sp., dated: 22.11.2006.

Habitat (Medicinal plants): Azadirachta indica, (Ghosh and Gupta, 2003), Zingiber sp. (new report)

Distribution: India: West Bengal (Gupta, 2002); Elsewhere: Mexico (Gonzalez-Rodriguez, 1965).

### 6. Agistemus unguiparvus Gonzalez-Rodriguez

1965. Agistemus unguiparvus Gonzalez-Rodriguez, Univ. Calif. Pub. Ent., 41-43.

Collection records: 3 females, India: West Bengal, Darjeeling, Sukna, ex *Aristolochia indica* L., dated: 24.11.2006.

Habitat (Medicinal plants): Citrus, cotton (Gupta, 2002), Aristolochia indica (new report).

Distribution: India: Uttar Pradesh, Tripura (Gupta, 2002), West Bengal (new report); Elsewhere: Mozambique (Gupta, 2002).

### Family 5. Tydeidae Kramer, 1877

### 7. Lorryia sp.

Collection records: 4 females, India: West Bengal, Darjeeling, Lloyed Botanical Garden, ex *Terminalia myriocarpa* Huerk & Muell., dated: 18.11.2006.

### Order II. Mesostigmata

### Family 6. Phytoseiidae Berlese, 1916

### 8. Amblyseius (Amblyseius) aerialis (Muma)

1955. Amblyseius (Amblyseius) aerialis (Muma), Ann. Ent. Soc. Amer., 48: 264-266.

Collection records: 9 females, India: West Bengal, Darjeeling, Lloyed Botanical Garden, ex *Clerodendron siphonanthus* R. Br., dated: 18.11.2006.

Habitat (Medicinal plants): Boerhavia diffusa, Carica papaya, Citrus sp. (Gupta, 2003, 2005). Clerodendron siphonanthus (new report).

Distribution: India: Arunachal Pradesh, Bihar, Karnataka (Gupta, 2003), West Bengal (new report); Elsewhere: U.S.A., Galapagos Isls., Mexico, Hondurus, Jamaica, Brazil, Algeria (Gupta, 2003).

### 9. Amblyseius (Amblyseius) herbicolus (Chant)

1959. Typhlodromus (Amblyseius) herbicolus Chant, Can. Ent., 91: 84-85.

Collection records: 15 females, 7 males, India: West Bengal, Darjeeling, Mungpoo, Govt. Quinine Factory Plantation area, ex *Cinchona officinalis* Linn., dated: 21.11.2006.

Habitat (Medicinal plants): Guava, papaya, wood apple, Hibiscus rosa-sinensis (Ghosh and Gupta, 2003). Aegle marmelos, Coccinia grandis, Ficus religiosa, Colocasia esculenta (Lahiri et al., 2004; Gupta, 2005)); Cinchona officinalis (new report).

Distribution: India: Arunachal Pradesh, Bihar, Tripura, West Bengal; Elsewhere: Philippines, Taiwan, Thailand, Japan, Madagascar, U.S.A., Mexico, Brazil, West Indies (Gupta, 2003).

### 10. Amblyseius (Amblyseius) largoensis (Muma)

1955. Amblyseiopsis largoensis Muma, Ann. Ent. Soc. Amer., 48: 266.

Collection records: 8 females, India: West Bengal, Darjeeling, Sukna, ex *Aristolochia indica* L., dated: 24.11.2006.

Habitat (Medicinal plants): Mangifera indica, Psidium guajava, Hibiscus rosasinensis (Ghosh and Gupta, 2003); Achyranthes aspera, Aegle marmelos, Anacardium occidentale, Azadirachta indica, Bauhinia purpurea. Bauhinia acuminate, Boerhavia diffusa, Carica papaya, Catharanthus roseus, Clerodendrum inerme, Coccinia grandis, Citrus spp., Curcuma aromatica, Datura metel, Ficus bengalensis, Ficus carica, Justicia adhatoda, Nerium indicum, Cassia fistula, Piper nigrum, Piper betle, Piper longum, Psidium guajava, Punica granatum, Rauvolfia tetraphylla, Ricinus communis, Thevetia nerifolia, Wissadula periplocifolia, (Lahiri et al., 2004; Gupta, 2005); Aristolochia indica (new report).

Distribution: India: Himachal Pradesh, Assam, Arunachal Pradesh, Bihar, Tripura, Manipur, West Bengal, Andhra Pradesh, Karnataka, Kerala, Andaman & Nicobar Isls., Gujrat; Elsewhere: Philippines, Taiwan, Thailand, Japan, Israel, Iran, South Africa, New Zealand, U.S.A., Mexico, Brazil, West Indies (Gupta, 2003).

### 11. Amblyseius (Euseius) coccineae Gupta

1975. Amblyseius (Euseius) coccineae Gupta, Indian J. Acar., 1: 30.

Collection records: 7 females, India: West Bengal, Darjeeling, 5th mile-Siliguri, ex *Morus alba* L., dated: 23.11.2006.

Habitat (Medicinal plants): Mangifera indica, Cocos nucifera, (Ghosh and Gupta, 2003), Anacardium occidentale, Carica papaya, Citrus spp., Coccinia grandis, Terminalia arjuna (Gupta, 2005); Morus alba (new report).

Distribution: India: Jammu& Kashmir, Uttar Pradesh, Bihar, Orissa, West Bengal, Meghalaya Andhra Pradesh, Karnataka, Kerala, Andaman & Nicobar Isls., Gujrat (Gupta, 2003).

### 12. Amblyseius (Euseius) coccosocius Ghai & Menon

1967. Amblyseius (Euseius) coccosocius Ghai & Menon. Oriental Ins., 1: 67-68.

Collection records: 2 females, India: West Bengal, Darjeeling, Salugara, Siliguri, ex *Desmodium motorium* (Houtt.) Merril., dated: 22.11.2006.

Habitat (Medicinal plants): Anacardium occidentale, Carica papaya, Citrus spp., Coccinia grandis, Terminalia arjuna (Gupta, 2005); Desmodium motorium (new report).

Distribution: India: Punjab, Tripura, West Bengal, Andhra Pradesh, Pondicherry, Tamil Nadu, Kerala, Karnataka, Lakshadweep (Gupta, 2003).

### 13. Amblyseius (Euseius) finlandicus (Oudemans)

1915. Seiulus finlandicus Oudemans, Ent. Ber., 4: 183.

Collection records: 11 females, 2males, India: West Bengal, Darjeeling, Sukna, ex *Quercus incana* R., dated: 24.11.2006.

Habitat (Medicinal plants): Carica papaya, Clitoria ternatea, Justicia adhatoda, Psidium guajava (Lahiri et al., 2004; Gupta, 2005)); Quercus incana (new report).

Distribution: India: Jammu& Kashmir, Punjab, Himachal Pradesh, Uttar Pradesh, Bihar, Tripura, Mizoram, Sikkim, Meghalaya, West Bengal, Karnataka, Lakshadwip Isls.; Elsewhere: Pakistan, Europe, Canada, Mexico, U.S.A., Africa, South America, Russia, Japan, Indonesia (Gupta, 2003).

### 14. Amblyseius (Euseius) pruni Gupta

1970. Amblyseius pruni Gupta, Internat. J. Acarol., 1(2): 40-42.

Collection records: 8 females, India: West Bengal, Darjeeling, Lloyed Botanical Garden, ex Clematis buchananian DC. dated: 18.11.2006.

Habitat (Medicinal plants): Psidium guajava, Glycosmis pentaphylla (Ghosh and Gupta, 2003); Bauhinia acuminate, Carica papaya, Citrus spp. (Gupta, 2005), Clematis buchananian (new report).

Distribution: India: Jammu& Kashmir, Punjab, Himachal Pradesh, Uttar Pradesh, Bihar, Sikkim, Assam, Tripura, Mizoram, Arunachal Pradesh, Meghalaya, West Bengal, Lakshadweep Islands. (Gupta, 2003).

### 15. Amblyseius (Typhlodromips) suknaensis Gupta

1970. Amblyseius suknaensis Gupta, Oriental Ins., 4: 185-186.

Collection records: 21 females, 9 males, India: West Bengal, Darjeeling, Salugara, Siliguri, ex *Barleria lupulina* Lindl., dated: 22.11.2006.

Habitat (Medicinal plants): Azadirachta indica, Citrus spp., Glycosmis pentaphylla (Ghosh and Gupta, 2003); Hibiscus abelmoschus, Abelmoschus moschatus, Catharanthus roseus, Datura metel, Piper betle (Lahiri et al., 2004; Gupta, 2005), Barleria lupulina (new report).

Distribution: India: Assam, West Bengal, Kerala, Andaman & Nicobar Isls. (Gupta, 2003).

### 16. Indoseiulus sp.

Collection records: 1 female, India: West Bengal, Darjeeling, Sukna, ex *Aristolochia indica* L., dated: 24.11.2006.

### 17. Phytoseius (Phytoseius) maldahensis Gupta

1992. Phytoseius (Phytoseius) maldahensis Gupta, In: State Fauna Ser. 3, Fauna of West Bengal, Part 3, p. 177.

Collection records: 4 females, India: West Bengal, Darjeeling, Salugara, Siliguri, ex Zingiber sp., dated: 22.11.2006.

Habitat (Medicinal plants): Mangifera indica (Gupta, 2003), Zingiber sp. (new report).

Distribution: India: West Bengal (Gupta, 2003).

### 18. Typhlodromus (Amblydromella) himalayensis Gupta

1981. Typhlodromus himalayensis Gupta, Indian J. Acar., 5(1-2): 32-33.

Collection records: 4 females, India: West Bengal, Darjeeling, Lloyed Botanical Garden, ex *Clematis buchananiana* DC, dated: 18.11.2006.

Habitat (Medicinal plants): Nerium sp. (Gupta, 2003), Clematis buchananiana (new report).

Distribution: India: Jammu& Kashmir, Himachal Pradesh, Uttar Pradesh (Gupta, 2003), West Bengal (new report) Andaman & Nicobar Isls., Lakshadweep Isl. (Gupta, 2003).

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#### REFERENCES

- Ghosh, S. and Gupta, S. K. (2003) A report on mites occurring on medicinal plants in West Bengal. *Records of Zoological Survey of India* 101: 287–298.
- Gonzalez-Rodriguez, R. H. (1965) A taxonomic study of the genera Mediotala, Zetzellia and Agistemus (Acarina: Stigmaeidae). *University of California Publications in Entomology* **41**: 64.
- Gupta, S. K. (2002) A Monograph on plant inhabiting predatory mites of India. Part I: Orders. Prostigmata, Astigmata and Cryptostigmata. *Memoirs of the Zoological Survey of India* 19(2): 183.
- Gupta, S. K. (2003) A Monograph on plant inhabiting predatory mites of India, Part II: Order Mesostigmata. *Memoirs of the Zoological Survey of India* 20(1): 185.
- Gupta, S. K. (2005) *Insects and Mites Infesting Medicinal Plants in India*, Ramakrishna Mission Ashrama, Narendrapur, Kolkata, 214.
- Haitlinger, R. (2004) New records of mites (Acari: Prostigmata: Erythraeidae, Trombidiidae, Eutrombidiidae) from Croatia, with descriptions of three new species. *Natura croatica* **13(2)**: 143–160.
- Khot, N. S. (1963) Studies on Indian Erythroidea (Acarina). Acarologia 5(2): 232-243.
- Lahiri, S., Podder, S., Saha, G. K. and Gupta, S. K. (2004) Diversity of phytophagous and predatory mites occurring on medicinal plants in Kolkata metropolis. *Proceedings of the Zoological Society, Kolkata* 57(1): 47–52.
- Slone, D. H. and Croft, B. A. (2001) Species association among predaceous and phytophagous apple mites (Acari: Eriophydae, Phytoseiidae, Stigmaeidae, Tetranychidae). *Experimental and Applied Acarology* **25**: 109–126.

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# A new genus and new species of Chloropidae (Diptera) from India

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ABSTRACT: A new genus *Paraapallates* with type species *P. convexa*, sp. n. is described from India. © 2008 Association for Advancement of Entomology

KEYWORDS: Diptera, Chloropidae, Paraapallates, gen. n., P. convexa, sp. n., India

### INTRODUCTION

Oscinellinae is the largest of the four subfamilies of Chloropidae. Flies of this subfamily are distributed in all the faunal Regions of the world. Of the 10 tribes recognized by Nartshuk (1987) under Oscinellinae, Incertellini Nartshuk (1983) is a medium sized one represented by six genera, namely *Apallates* Sabrosky, *Aphanotrigonum* Duda, *Biorbitella* Sabrosky, *Malloewia* Sabrosky, *Incertella* Sabrosky and *Meijerella* Sabrosky. Of these five genera were erected by Sabrosky (1980) and only the last two have been reported from India.

During a revisionary study of Chloropidae of India and adjacent countries, we came across a species from Kerala in Southern India that does not fit elsewhere in the classification of the family though it shows affinities to *Apallates* belonging to tribe Incertellini. The species is placed under a new genus, *Paraapallates* with type species *P. convexa*, sp. n. Both the new genus and species are described here.

Type specimens are retained in the collections of the University of Kerala and shall later be deposited in the National Collections of Zoological Survey of India, Kolkata.

### Paraapallates Cherian, gen. n.

Type species: Paraapallates convexa Cherian, sp. n.

Small, partly yellow flies with tomentose frontal triangle, well developed body bristles, evenly distributed scutal hairs, brownish black haltere and short preapical, anteroventral spur on hind tibia.

Head: Higher than long with dark hairs and bristles; from nearly parallel sided, wider

at vertex than long with fairly dense, conspicuous fr; frontal triangle not very distinctly demarcated, apparently reaching three-fourths length of frons, finely tomentose; face deeply concave; facial carina fades off before middle of face. Antenna erect; ant<sub>3</sub> subreniform, wider than long; arista slender, brownish black with conspicuous pubescence. Gena about half as wide as ant<sub>3</sub>; vibrissal corner receding; postgena reduced; parafacialia not developed. Eye large, sparsely and finely pubescent, with vertical long axis. Head bristles well developed; ovt longer than ivt; pvt convergent; oc reclinate; orb 4, reclinate; if just outside margin of frontal triangle and posteriorly a few along its margin.

Thorax: As wide as head. Scutum convex, densely tomentose, yellowish brown with three broad partly diffused brownish black longitudinal bands and evenly distributed short hairs. Pleura predominantly black; anepst and anepm tomentose and rest of the areas polished. Scutellum nearly subtriangular, wider than long with slightly convex disc which appears nearly flattened in certain angles of illumination. Thoracic bristles black, very well developed with one h, 1 + 2 npl, 1 dc and pa 1; pa 2 not developed; as much longer than scutellum; ss 1 more than half as long as as.

Wing: Hyaline with pale brown veins; costa reaching m 1+2 which ends at apex of wing; length of second costal sector more than combined length of third and fourth sectors; discal cell of *Oscinella*-type. Anal area slightly receding. Haltere brownish black.

Legs: Slender, hind tibia with short, black, preapical anteroventral spur which is shorter than diameter of tibia; femoral organ in the form of a group of short warts nearly in two rows; tibial organ long and well developed.

Abdomen: Almost as wide as thorax, finely dark tomentose with a few pale hairs. Female cerci of medium size. Male genitalia: Cerci large, widely separated, distally becoming narrow along one-third their lengths, surstylus broad and flat basally, gradually narrowing distally and ending with obtuse apex; hypandrium complete, aedeagal apodeme projecting posteriorly far above hypandrium; postgonite long and well developed.

Distribution

Oriental Region.

Gender and derivation

Masculine based on genus Apallates with the prefix para.

### Remarks

In the development of a preapical, anteroventrally located spur on hind tibia, genera *Apallates* and *Paraapallates* show similarity. But while in the former, frontal triangle is chiefly polished black, face is weakly concave, cephalic bristles are slender and not very conspicuous, there are 7 orb, scutal hairs are arranged in rows, scutellum is broadly rounded and haltere is yellow, in *Paraapallates* frontal triangle is finely

but distinctly tomentose, face is deeply concave, cephalic bristles are well developed, there are only 4 *orb*, scutal hairs are uniformly distributed and not in rows, scutellum is nearly subtriangular and haltere is brownish black. Besides, there are marked differences in the male genitalia of the two genera. In the tomentose nature of frontal triangle, uniformly distributed scutal hairs, nature of wing veins and such other characters, *Paraapallates* resembles some species of *Conioscinella*. However, apart from other characters including differences in the male genitalia, in *Paraapallates* haltere is brownish black and there is a distinct preapical spur on hind tibia whereas in *Conioscinella* haltere is yellow and preapical hind tibial spur is absent.

### Paraapallates convexa Cherian sp. n. (figures 1-5)

### Male and Female

Head (Fig. 1): Mostly brownish yellow, length, height and width ratio 6:7:9. Frons brownish yellow, nearly parallel sided but widening at vertex, width at point of widening at vertex 0.6 x that of head and equal to its own length, slightly sloping at sides anteriorly, ending with convex margin which does not reach anterior margin of eye and with well developed brownish black fr of which a pair medially just behind anterior margin more developed, partly proclinate and convergent at tips; frontal triangle not clearly demarcated, apparently appearing to reach three-fourths length of frons, finely grey tomentose; if in a row along margin of triangle and a few posterior ones on the triangle within its margin. Face deeply concave, finely grey tomentose; facial carina triangular between bases of antennae and reaching linearly almost middle of face whence it fades off; epistomal margin not thickened, distinctly raised. Antennae erect; basal antennal segments yellow; ant<sub>3</sub> subreniform, about 1.4 x as wide as long, upper half brownish black, lower half brownish yellow; arista thickened at base, gradually narrowing and becoming very slender distally, brownish black with distinct concolorous pubescence. Gena a little less than half as wide as ant3, finely grey tomentose with a row of oral setae; vibrissal corner distinctly receding, not reaching anterior margin of eye; postgena a little reduced, concolorous with gena. Eye large, sparsely and finely pubescent, with vertical long axis. Parafacialia not developed. Palpi rather slender, cylindrical, yellow; proboscis short with deep brownish tinge. Head bristles well developed, brownish black; ivt shorter than ovt, the latter subequal to convergent pvt; oc reclinate, distally convergent; orb 4, very well developed, reclinate; oral vibrissa conspicuously developed.

Thorax: Scutum prominently convex, fairly densely grey tomentose, yellowish brown with three broad brownish black partly diffused longitudinal bands of which median is longitudinally faintly subdivided and each lateral one is interrupted at transverse suture and all bands posteriorly abbreviated at around level of 1 dc; scutal hairs short, evenly distributed, yellow; pleura as described under the genus. Scutellum (Fig. 2) subtriangular, two-thirds as long as wide with gently convex disc which appears nearly flattened in some angles of illumination, partly grey tomentose, yellowish brown with dark tinge and a few scattered yellow hairs. Thoracic bristles well developed, with

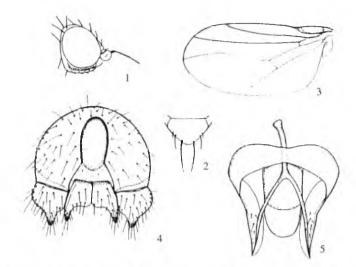


FIGURE 1. Paraapallates convexa Cherian, sp. n. Head; 2. Scutellum; 3. Wing; 4&5. Male genitalia.

1h, 1 + 2 npl and subequal pa1 and 1 dc; as 1.2 x as long as scutellum, fairly widely separated at base, becoming divergent distally; ss 1 about half as long as as and widely separated from base of as; ss 2, 0.4 x as long as ss 1.

Wing (Fig. 3): 2.3 x as long as wide, hyaline with deeply brown veins; proportions of costal sectors 2 to 4 in the ratio 17:8:6; terminal sector of r 4+5 nearly straight, that of m 1+2 conspicuously convex above medially, recalling the condition in *Conioscinella semimaculata* (Becker); discal cell a little narrowed; r-m cross vein distad of middle of discal cell, opposite 0.58 of its length; anal area slightly receding. Haltere brownish black.

Legs: Slender, brownish yellow but for slightly infuscated fore tarsi, last tarsal segment of hind leg and diffused large infuscation on midfemur, tibia and tarsi of one side and diffused dark tinge on hind femur in the holotype, and almost wholly infuscated foreleg and mid and hindfemora of one side and light infuscation on mid and hind femora on other side in the paratype. As infuscation on legs of both the holotype and paratype are not uniform and differs in legs on two sides of the same specimen, it is probable that it was partly contributed by the mode of collection and nature of preservation. Tibial and femoral organs as described under the genus; anteroventral hind tibial spur a little shorter than diameter of hind tibia.

Abdomen: Dull black, finely and darkly tomentose, fairly long with slender, pale hairs especially on distal segments. Female cerci of medium size with a few slender hairs distally. Male genitalia (Fig. 4 and 5): Cerci large, basally the two meeting in the middle, distally widely separated, becoming narrow along one-third their lengths; surstylus broad and flat basally, gradually narrowing along one-third its length

distally and ending with obtuse apex bearing long hairs especially on outer serrated margin; hypandrium complete; aedeagal apodeme projecting far above hypandrium posteriorly; postgonite well developed, gradually narrowing and becoming pointed terminally.

Length

Male: 1.5 mm; wing: 1.52 mm Female: 1.7 mm; wing: 1.85 mm.

Holoytpe

Male, India: Tamil Nadu: Nagerkovil, 24.iii.1988, Coll. Koshey Mathew.

Paratype: 1 female, India, Tamil Nadu: Kanyakumari Dt., Kodayar, 30.iii.1988, Coll. Koshey Mathew.

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### **ABBREVIATIONS**

anepst – anepisternum; anepm – anepimeron; ant<sub>3</sub> – third antennal segment; as – apical scutellar bristle; dc – first dorsocentral bristle; fr – frontal hair; fr – humeral bristle; fr – interfrontal bristle; fr – inner vertical bristle; fr – notopleural bristle; fr – ocellar bristle; fr – fronto – orbital bristle; fr – outer vertical bristle; fr – first postalar bristle; fr – second postalar bristle; fr – postvertical bristle; fr – subapical scutellar bristle.

### REFERENCES

- Andersson, H. (1977) Taxonomic and phylogenetic studies on chloropidae (Diptera) with special reference to Old World genera. *Ent. Scand. Suppl.* 8: 1–200.
- Cherian, P. T. (1989) Some new genera of oriental chloropidae (diptera). *Orient. Insects* 23: 219–229.14 figures
- Cherian, P. T. (2002) Fauna of India and the Adjacent Countries—Diptera Vol.IX Chloropidae (Part 1), Published-Director, ZSI, Kolkata, 1–368.
- Kanmiya, K. (1983) A systematic study of the Japanese Chloropidae (Diptera). *Mem. Ent. Soc. Washington* 11: 132–134.
- Nartshuk, E. P. (1983) A system of the superfamily chloropoidea (diptera: cyclorrhapha). *Ent. Oboz.* **62(3)**: 638–648.
- Nartshuk, E. P. (1987) Zlakovie Mukhi (Diptera: Chloropidae) IK Systema, Evolusia i Suyazi s rastennymi. *Trud. Zool. Inst. Acad. Nauk USSR* **136**: 1–280.
- Sabrosky, C. W. (1980) New genera and new combinations in Nearctic Chloropidae (Diptera). *Proc. Ent. Soc. Was.* **82(3)**: 412–429.

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# Comparative genotoxicity of alpha-cyano pyrethroids on *Drosophila melanogaster*

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ABSTRACT: The polytene chromosomes found in third instar larvae of *Drosophila melanogaster* consist of puffs in which DNA strand uncoil, protrude out as a loop and produces number of copies of mRNA and dark and light bands which contain various concentrations of DNA and proteins in the chromatin. Number of puffs and dark bands increased after intoxication with cypermethrin and alphamethrin. © 2008 Association for Advancement of Entomology

KEYWORDS: Drosophila melanogaster, Polytene chromosome, effect of cypermethrin, alphamethrin treatment

The salivary glands of *Drosophila melanogaster* third instar larvae have polytenized interphase chromosome and are an excellent experimental system in which puffing pattern and hence gene activity can be demonstrated (Dworniczak *et al.*, 1983). Normally during interphase the chromosome would not be visible but these polytene chromosomes consist of many duplicated strands of DNA. This structure is a type of gene amplification which allows rapid protein synthesis. The banding pattern represents areas of gene activity and inactivity.

It is well established that even very low doses of pyrethroid insecticides are able to induce marked increase in the frequency of electrical activity of insect neurosecretory cells, which may result in the release of neuro-hormones. Beside this many physiological and biochemical effects have been described in tissues outside the nervous system (Saleem et al., 1998; Saleem and Shakoori, 1996, 1987). Very few reports have appeared on the genotoxicity of synthetic pyrethroids. The present study is expected to produce insight into the effect of sub-lethal doses of cypermethrin and alphamethrin on polytene chromosome of white and sepia mutants of *Drosophila melanogaster* 

<sup>\*</sup>Corresponding author

TABLE 1.							and	white
	mutant	s of <i>Dro</i>	sophi	la melan	oge	aster		_
						****		

	Sep	ia mutant	Whit	e mutant
Sets	No. of puffs	No. of dark bands	No. of puffs	No. of dark bands
Control	7	87	17	100
Alphamethrin treated	24	140	28	142
Cypermethrin treated	21	105	21	134

Pure culture of sepia and white mutants of *Drosophila melanogaster* was maintained at Department of Zoology, SLS, Khandari, Agra. The culture was nurtured and maintained in glass culture vials of 100 ml capacity, at a temperature 25 °C, 50% relative humidity in B.O.D. incubator. The flies were fed on prescribed Drosophila food (Roberts 1986). Median Lethal concentration (LC<sub>50</sub>) of the two insecticides were assessed following bioassay procedures in the laboratory and subjecting the mortality data to probit analysis (Finney 1971.) Polytene chromosome preparations were made by the conventional squash procedure using 2% aceto-carmine. All squashes with well spread polytene chromosome were processed for permanent slides which were photographed with the help of Motic Microscope system at 1000X.

Based an L.C 50 of the pesticides, cypermethrin and alphamethrin were sprayed on D. melanogaster flies taken in petridishes. Unsprayed lots were kept as control. At the end of 48 h after treatment flies in an all treatments (including control) were allowed to mate for five days and the experiment was conducted in  $F_1$  generation. Third instar larvae of this generation were used for making the chromosome preparations.

The number of puffs and dark bands were found increased after treatment with  $\alpha$ -cyano pyrethroids, cypermethrin as well as alphamethrin (Table 1, figure 1a, b and c). Therefore it could be inferred that experimental compounds not only affect nervous system, biochemistry and physiology but also genetic makeup of organism with increase in number of puffs and dark bands. DNA, RNA and protein also increased as it has already been mentioned that puffs contain mRNA and dark bands made of DNA and protein. Alphamethrin was found more potent than cypermethrin and white mutants were more susceptible than sepia mutant in the present investigation.

By correlating puffs with different physiological or developmental process, scientists have been able to locate genes on the polytene chromosome and prepare chromosome maps.

Normally gene activation in dipteran polytene chromosome is accompanied by a local decondensation of the bands and puffing. Three hundred and fifty puffed regions that have been described in *Drosophila melanogaster* salivary gland polytene chromosome to date are either light (or smooth) and relatively homogenous, or dark, with a more dense sometimes granular structure (Semestin *et al.*, 2001). Bands in such regions are decondensed incompletely, their diameters slightly exceed those of the neighbouring dense bands (Semestin *et al.*, 2001).

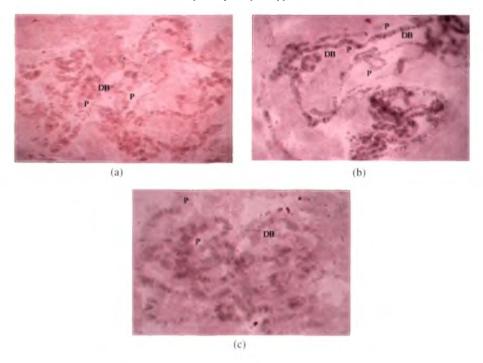


FIGURE 1. (a) Puffs (P) and dark bands (DB) in polytene chromosomes of sepia mutant of *Drosophila melanogaster* (control set) [1000X] (b) Increase in the number of puffs (P) and dark bands (DB) in polytene chromosomes of cypermethrin treated sepia mutant of *Drosophila melanogaster* [1000X] (c) Increase in the number of puffs (P) and dark bands (DB) in polytene chromosomes of alphamethrin treated sepia mutant of *Drosophila melanogaster* [1000X]

The extent of protein induction and the degree of puff induction are related to the severity of the treatment. Similar to the present findings induction in puffs within 30–40 min after incubation with caffeine that attained maximum size after 60 min in *Drosophila* melanogaster was observed (Srivastava and Bangia, 1985); hexoses as well as some disaccharides, except lactose, as efficient inducer and alcohols, such as ethanol and glycerol, showed a moderate capacity as inducers, however, mixture of galactose and glycerol was found to be significant accelerator at the puffing as well as translational level in *Chironomus thummilaria* (Cortes *et al.*, 1990) and chromosomal aberration as synapsis, ectopic pairing and chromosome break in the hydrogen peroxide treated polytene chromosome of *Chironomus samoensis* Edwards was reported (Khanna *et al.*, 2006).

White mutant flies have been found to be more susceptible to experimental compounds as compared to the sepia. This differential response may be attributed to the genetic makeup of these mutants. Toxicological and pharmacological data shows that resistance involves a modification of the affinity of the insecticide for its receptor site on the voltage dependent sodium channel (Amichot *et al.*, 1992). Genetic studies

138 N. Rana *et al*.

indicate that the kdr factor is linked to the second chromosome where one sodium channel gene sch is located. Cloning and sequencing of this gene from resistance and susceptible strains revealed a single substitution that may be responsible for the loss of toxicity of insecticide in the resistant strains (Nadda *et al.*, 2005).

It could be concluded from the present investigation that alphamethrin is more potent genotoxic compound in comparison to cypermethrin. It might be because alphamethrin contains two active isomers (more than 90%) of the four cis isomers of cypermethin.

### REFERENCES

- Amichot, M., Castella, C., Cuany, A., Berge, J. B. and Pauron, D. (1992) Target modification as a molecular mechanism of pyrethroid resistance in *Drosophila melanogaster*. *Pest. Biochem. Physiol.* **44(3)**: 183–190.
- Cortes, E., Seerano, M., Yanez, R. J. and Diez, J. L. (1990) Induction of Balbiani during puffing changes by sugars and alcohols in *Chironomus thummi*. *Ins. Biochem.* **20(5)**: 523–529.
- Dworniczak, B., Seidel, R. and Pongs, O. (1983) Puffing activities and binding of ecdysteroid to polytene chromosome of *Drosophila melanogaster*. The Embo J. **2**(3): 1323–1330.
- Finney, D. J. (1971) Probit Analysis, Cambridge University Press, 303.
- Khanna, P., Sharma, O. P. and Tripathy, N. K. (2006) Genotoxic effect of hydrogen peroxide on polytene chromosomes of *Chironomus samoensis* Edwards (Chironomidae: Diptera). *Proc. Nat. Acad. Sci. India* **76(B)**, **1**: 41–50.
- Nadda, G., Saxena, P. N. and Śrivastava, G. (2005) Effects of beta-cyfluthrin on white and sepia mutants of *Drosophila melanogaster*. *J. Environ. Biol.* **26(2 suppl.)**: 363–367.
- Roberts, D. B. (1986) *Drosophila, A practical approach*, IRL Press Oxford, OX8, IJJ, England, 295
- Saleem, M. A. and Shakoori, A. R. (1987) Permethrin and malathion induced macromolecular abnormalities in adults of *Tribolium castaneum* (Herbst.). *Arch. Insect. Biochem. Physiol.* 5: 45–57.
- Saleem, M. A. and Shakoori, A. R. (1996) Biochemical studies on Talcord 10EC. II. Effect on some enzyme activities and macromolecules of adult beetles of *Tribolium castaneum*. *Pak. J. Zool.* 28(2): 151–162.
- Saleem, M. A., Shakoori, A. R. and Mantle, D. (1998) In vivo ripcord induced macromolecular abnormalities in *Tribolium castaneum* larvae. Pak. J. Zool. 30(3): 233–243.
- Semestin, Y. F., Shloma, V. V. and Zhimulev, 1. F. (2001) Formation and morphology of dark puffs in *Drosophila melanogaster* polytene chromosomes. *Heriditas* 134: 15–22.
- Srivastava, J. P. and Bangia, K. K. (1985) Effect of caffeine on puffing pattern in the polytene chromosomes of salivary glands of *Drosophila melanogaster*. *Cur. Sci.* **54(13)**: 651–652.

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### Effect of age of the host plant on the performance of Ceutorhynchus portulacae Marshal (Coleoptera: Curculionidae), a herbivore of Portulaca oleracea

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ABSTRACT: Performance of the specialist herbivore *Ceutorhynchus portulacae* was studied in relation to age of its host plant *Portulaca oleracae*. Significant reduction in longevity and fecundity of adults were recorded in relation to age of the host plant provided. Advance in the age of the host plant was found to affect the growth, development and fecundity of the insect. The physical characteristics and chemical composition of host plant also influenced the establishment of the insect. The implications of the results in relation to biological control of purslane weed *P.oleracae* are discussed. © 2008 Association for Advancement of Entomology

KEYWORDS: *Portulaca oleracea*, *Ceutorhynchus portulacae*, biotic and functional potential, host plant phenology

Purslane weed *Portulaca oleracea* L. (Portulacaceae) ranks as one of the world's worst weeds and is of considerable importance in many agricultural crops of tropical countries (Holms *et al.*, 1977). The weevil, *Ceutorhynchus portulacae* Marshall (Coleoptera: Curculionidae) was identified as a potential natural enemy of the weed that could be effective in biological suppression of the weed (Ganga Visalakshy and Jayanth, 1995). *C. portulacae* larvae mines leaves, stem and seed capsules. Adult weevils feed on leaves, tender stems and on seed capsules. The feeding results in skeletonizing and drying up of entire plant (Ganga Visalakshy, 2007). Aim of the present study was to determine influence of host plant age on survival, growth, development, feeding preference, fecundity and longevity of larvae and adults and relate it with chemical and physical characteristics of plant.

Purslane plants were grown in pots under glass house conditions. From these 20–30 days and 45–60 days old plants were selected for study. To determine rate of oviposition bouquets of twigs of above-mentioned ages were kept separately in oviposition cage (The oviposition cage was a plastic jar  $15 \times 10$  cm with an aerated lid at the top.) Purslane twigs of 10 cm length were cut from plants maintained in glass house; bouquets were made by wrapping the base of twigs in wet cotton held

together by a rubber band. The cotton swab was moistened as and when needed to prevent twigs from drying. Twenty four hour old adults were released into oviposition cage. Bouquets were replaced daily with fresh ones. Eggs laid on exposed leaves were recorded. The experiment was continued till all the exposed adults were dead. The feeding marks on the leaves exposed to the weevils for oviposition was also recorded till the adults died. The total number of feeding marks was divided by the longevity of the adults to obtain the feeding per day.

To determine effect of plant age on larval growth and survival, newly emerged larvae were placed in ventilated plastic jars as mentioned above ( $15 \times 10$  cm) containing bouquets from purslane plants of different ages. Fresh bouquets of similar age were provided as and when required. Larvae after completing development drops to the base of jar for pupation. The number of larvae that completed development, their weight and percent survival were recorded. The experiment was replicated five times with ten larvae per replication.

Concentrations of major elements in leaf samples were estimated drying them in oven at 80 °C for 72 hrs and powdering them in a wiley mill. One gram of the sample was digested using perchloric acid, nitric acid in the ratio of 9:2. Digested acid samples were used for estimating nutrient in a flame photometer/colorimeter. Concentration of nutrient is expressed on a dry matter basis. Leaf thickness was measured in cross sections under a microscope by using an ocular meter.

Plant age affected the biological parameters of *C. portulacae* larvae and adult. A greater percentage of larva survived on younger leaves (Table 1). However the development duration and weight gained by larvae in the two treatments were similar. When adults were exposed to leaves of different ages of purslane, area of leaf fed, longevity and fecundity were seen reduced in older leaves compared to younger leaves. Differences in external texture, colour, and thickness and chemical composition of the leaves of different ages were recorded. Leaves of young plants were soft to touch and light green in color, while those of older plants were dark green and hard. Similarly, younger leaves were less thick than older (Table 2). Zinc (Zn) was completely absent in younger plants while it was high in older leaves. Copper (Cu) was eight times less in older plants, while sodium was four times less in older plants (Table 2). Calcium (Ca) content was 50 per cent more in older plants than in younger plants. Potassium (P) and Manganese (Mn) concentrations were similar in leaves of different ages.

Results of the present study suggest that physical and chemical characteristics of purslane plants vary with plant age and that these changes affect growth, survival, feeding preference, fecundity and longevity of *C. portulacae* larvae and adults. The importance of physical and chemical factors for successful biocontrol programmes has been documented for alligator weed (Maddox and Rhyne, 1975), *Slavonia molest* (Taylor, 1984), *Eichhornia crassipes* (Center and Wheeler, 1991), Hydrilla (Wheeler and Center, 1996), and *Parthenium hystereophorus* (Annadurai, 1990).

TABLE 1. Effect of leaf age of purslane weed on the development of C. portulacae

Parameters	Age of th	e plant
studied	20-30d days	45-60d old
Duration (days)	6.10	6.30
Mortality (%)	10.65*	52.00
Larval weight (g)	1.73	1.26
No. of feeding		
marks/day/adult	4.9*	2.5
Eggs/female/day	6.7*	1.1
Longevity (days)	69.7*	39.4

<sup>\*</sup>Significant at 0.05%

TABLE 2. Effect of age on leaf thickness and concentration of major elements in leaves of *P. oleracae* 

Parameters	Age of th	ne plant
studied	20-30d old	45–60d old
Leaf thickness (mm)	0.34	0.74
Na content (ppm)	1.30	0.30
P content (ppm)	2.03	2.03
Mn content (ppm)	59.00	52.00
Zn content (ppm)	0.00	96.00
Cu content (ppm)	120.00	15.00
Ca content (ppm)	2.43	4.01

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### REFERENCES

Annadurai, R. S. (1990) Reproductive potential in term of quantitative food utilization of *Zygogramma bicolorata* on *Parthenium hysterophorus*. In: *Proc. of VIIIth Internat. Sym. on Biol.Control of Weeds*, Insituto sperimentaleper laPata Rogia Vegetable Ministero delle Foresta, Rome, Italy, 385–394.

Center, T. D. and Wheeler, G. S. (1991) Age and phytochemical composition of water hyacinth leaves determined their acceptability to *Neochetina* spp. *Environmental Entomol*. 20: 323–324.

Ganga Visalakshy, P. N. and Jayanth, K. P. (1995) Ceuthorhynchus portulacae, Marshall (Coleoptera: Curculionidae) a potential biological control agent of Portulaca oleracea. Entomon 22: 150–151.

- Ganga Visalakshy, P. N. (2007) Biological studies on *Ceutorhynchus portulacae*, a potential natural enemy of the purslane weed, *Portulaca oleracea*, *Biocontrol* **52**: 619–628.
- Holms, L. G., Plucknett, D. L., Pancho, J. V. and Herberger, J. P. (1977) The World's Worst Weeds, *Distribution and Biology*., Honolulu HI University Press of Hawaii.
- Maddox, D. M. and Rhyne, M. (1975) Effect of induced host plant mineral deficiencies on attraction, feeding and fecundity of the alligator flea beetle. *Environmental Entomology* 4: 682–686.
- Taylor, M. F. J. (1984) The dependence of development and fecundity of *Samea multiplicalis* on early larval nitrogen intake. *J. of Insect Physiology* **30**: 779–785.
- Wheeler, G. S. and Center, T. D. (1996) The influence of hydrilla leaf quality on larval growth and development of the biological control agent *Hydrilla pakistanae* (Diptera: Ephyelidae). *Biological Control* 7: 1–9.

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## Butea monosperma (Lam.), a new host of Lampides boeticus Linnaeus in Rajastan, India

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ABSTRACT: Extensive incidence of the lycaenid butterfly *Lampides boeticus* Linnaeus was observed as a pest damaging mainly the unopened flowers and occasionally the pods and seeds of *Butea monosperma* (Lam.) Taub 'the flame of the forest' in and around Udaipur, Rajasthan, India. This is the first record of the pest on *B. monosperma*. Brief observation on the life stages of the pest is also included. © 2008 Association for Advancement of Entomology

KEYWORDS: Lampides boeticus, Butea monosperma, new host record

Lampides boeticus Linnaeus is a highly polyphagous pest attacking a wide variety of cultivated and wild varieties of legumes. Commonwealth Agricultural Bureau has recently (2004) catalogued 22 plant species as primary hosts of *L. boeticus*, seven (broom, corkwood, etc) as secondary hosts and eight as wild species. Many earlier reports on the occurrence of the pest on a variety of plants and in different countries of the world are available in literature. These include *Ulex europaeus* in New Zealand (Harding, 1971), *Crotalaria cunninghami* in Australia (Common and Waterhouse, 1981) *Medicago sativa* in Spain (Martin Cano, 1984) *Peuraria phaseoloides* in Saudi Arabia (Pittaway, 1985), *Lablab purpureus* and *Phaseolus vulgaris* in Taiwan (Chang and Chen, 1989), *Brassica* sp. (Mavi, 1992) and *Vigna exillata* in India (Govindan *et al.*, 1989).

In 2006 extensive incidence of *L. boeticus* was observed on *Butea monosperma* (Lamb) Taub, ('flame of the forest') in Gogunda block area of Udaipur district of Rajasthan. The larvae were seen entering largely in the unopened flower buds and eating from within. Sometimes they enter the pods and seeds too. Being a new host, preliminary observations on the life stages of the pest on the host was recorded in the laboratory by keeping the flower buds with insect egg in 500 ml glass jars closed with muslin cloths held in position with rubber bands. Moths lay egg singly on unopened flowers, on the flower stalk or sepals. Eggs are around 0.34 mm length, toroidal and china white in colour (figure 1). First instar larva is citrine yellow and in second and third instar the colour turns into yellowish red. All stages have a purple brown median

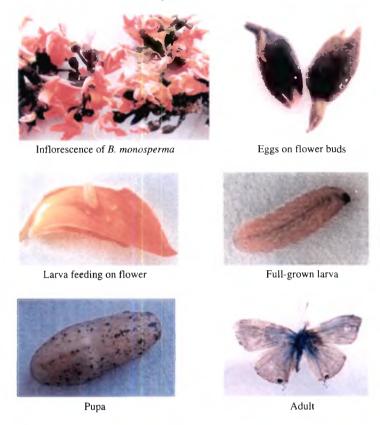


FIGURE 1. Life stages of *Lampides boeticus* (a) Eggs on flower buds; (b) Larva feeding on flower; (c) Full-grown larva; (d) Pupa; (e) Adult.

dorsal stripe, reddish lateral streak on either side, brown head, short marginal hairs and a dense covering of minute setae on body tubercles. Pupae have pale flush colour in the begining which later becomes pale brown. The pupa has a dorsal dark line and a varying number of brownish black spots. Wings of adult males are purplish-blue on the upper side and females possess wings having dark brown colour above.

### **REFERENCES**

CABI, (2004) Crop Protection Compendium, Global Module, 2004 edition, CAB International, Wallingford, United Kingdom.

Cameron, D. G. (1986) Tropical and sub-tropical pasture legumes. 12. Puero (*Pueraria phaseoloides*): a much underused legume. *Queensland Agricultural Journal* 112: 227–230.

Chang, T. C. and Chen, C. C. (1989) Observation of three lepidopterous pests attacking leguminous vegetables in Taiwan. *Bulletin of Taichung District Agricultural Improvement Station* **24**: 211–29.

Common, I. F. B. and Waterhouse, D. F. (1981) *Butterflies of Australia Review Edn*, Angus and Robertson Publishers, Melbourne, Australia.

- Govindan, R., Sarayanaswamy, T. K., Gururajarao, M. R. and Satenahalli, S. (1989) Insects infesting wild mung, *Vigna vexillata* in India. *Environmental and Ecology* 7: 513–513.
- Harding, J. W. (1971) Observations on Lampides boeticus (L.) (Lepidoptera: Lycaenidae). New Zealand Entomologist 5: 70–73.
- Howarth, T. G. (1973) South's British Butterflies, Frederick Warne & Co., London, U.K..
- Martin Cano, J. (1984) Comparative biology of *Lampides boeticus* (L.) *Syntarucus pirithous* (L.) and *Polyommatus incarus* (Rot.) (Lepidoptera: Lycaenidae). *Graellisia* **40**: 163–193.
- Mavi, G. S. (1992) A critical review on the distribution and host range of pea blue butterfly, Lampides boeticus (L.), Journal of Insect Sciences 5: 115-119.
- Pittaway, A. R. (1985) Lepidoptera: Rhopalocera of Western Saudi Arabia, Fauna of Saudi Arabia, 172–197.

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# Identification of promising multivoltine $\times$ bivoltine hybrids of the mulberry silkworm, *Bombyx mori* L.

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ABSTRACT: Thirty multivoltine  $\times$  bivoltine hybrids developed through application of androgenesis at Central Sericultural Research and Training Institute, Mysore were evaluated for eleven economic characters following subordinate function index method and multiple traits evaluation index method. Two hybrids AGL $_3 \times \text{CSR}_2$  and AGL $_5 \times \text{CSR}_2$  were found promising, exhibiting significant improvement in several quantitative characters, high cumulative index value (9.7 and 10.4) and average evaluation index value (65.25 and 68.52). They are recommended for large scale evaluation by the farmers. © 2008 Association for Advancement of Entomology

KEYWORDS:  $Bombyx\ mori$ , multivoltine  $\times$  bivoltine hybrids, multiple traits evaluation index, subordinate function index

Superiority of silkworm breeds and hybrids is judged on the basis of cumulative effect of several characters (Narayanaswamy et al., 2002). In the mulberry silkworm, Bombyx mori L., the multiple traits evaluation index method of Mano et al. (1993) and subordinate function index method of Gower (1971) have been employed simultaneously for the identification of promising silkworm hybrids (Ramesh Babu et al., 2002; Rao et al., 2001, 2004, 2006; Lakshmi and Chandrashekharaiah, 2007). In the present study, an attempt has been made to identify promising multivoltine × bivoltine hybrids developed utilizing androgenesis coupled with conventional breeding techniques.

In the present study, thirty multivoltine  $\times$  bivoltine hybrids were prepared utilizing six multivoltine breeds viz., AGL<sub>1</sub>, AGL<sub>2</sub>, AGL<sub>3</sub>, AGL<sub>4</sub>, AGL<sub>5</sub> and Pure Mysore (PM) and five bivoltine breeds viz., CSR<sub>2</sub>, CSR<sub>3</sub>, CSR<sub>4</sub>, CSR<sub>12</sub> and NB<sub>4</sub>D<sub>2</sub>. Three replications were reared for each hybrid and 250 larvae were retained after third moult in each replication. The performance of ten top ranking hybrids along with control (PM  $\times$  CSR<sub>2</sub>) is presented in Table 1.

Data presented in Table 1 show significant variation for various characters among the different hybrids. Two hybrids namely,  $AGL_3 \times CSR_2$  and  $AGL_5 \times CSR_2$  exhibited

<sup>\*</sup>Corresponding author

TABLE 1. Performance of selected multivoltine × bivoltine hybrids

					1								
Hybrids	Fecundity (No.)	Hatching (%)	Pupation Rate (%)	Yield/10,000 wt. (kg)	Coccoon wt (g)	Cocoon shell wt	Cocoon shell (%)	Filament length (m)	Reel- ability (%)	Raw silk (%)	Neat- ness (p)	Cumulative	Average Evaluation Index
AGLs × CSR3		0.96	97.20	69.61	2.041	0.438	21.44	885	84.33	15.53	7.06	10.40	68.52
AGL3 × CSR3		96.2	09'96	18 32	1.928	0.412	21.38	870	82.33	16.17	7.06	67.6	65.25
AGL, x CSR1,		94.6	97.33	16.97	1.775	0.377	21.22	808	79.00	15.10	7.68	7.47	56.18
AGL4 × CSR2		6.96	09.76	16.98	1.782	0.365	20.50	808	80.00	14.33	0.06	7.44	56.03
AGL, × CSR		96.3	94.80	16.43	1.756	0.359	20.46	814	81.33	14.47	90.3	7.43	49.07
AGL <sub>2</sub> × CSR <sub>2</sub>		95.4	97.47	16.86	1.764	0.364	20.62	992	79.33	15.40	88.3	7.30	50.16
AGL, x CSR,		95.3	97.73	17.04	1.782	0.369	20.70	781	78.33	16.60	89.3	7.29	55.45
AGLS × CSR12		95.5	96.27	16.60	1.749	0.367	21.01	786	80.00	15.70	89.7	7.27	55.23
AGL <sub>1</sub> × CSR <sub>2</sub>		94.0	95.20	17,42	1.866	0.371	19.88	811	79.00	14.90	7.68	7.15	55.24
AGL, × CSR3		9.96	97.73	16.84	1.755	0.354	20.15	727	79.00	14.63	7.68	7.05	54.21
PM × CSR <sub>2</sub>	422	94.5	09.76	16.92	1.762	0.336	19.05	778	29.08	13.73	87.0	5.47	48.88
(Control)													
CD@5%		1.4	,	0.52	0.055	0.013	0.50	50	ľ	1.00	=	ľ	Į.
CD @ 1 %	29	1.9	ì	0.71	0.075	0.018	0.67	89	)	1.36	1.5	Ŷ	.1

significantly higher values for most of the economic characters. It was interesting to note that all the ten top ranking hybrids have shown significantly higher values for three characters namely, cocoon shell weight, cocoon shell percentage and neatness. The cumulative index was highest in  $AGL_5 \times CSR_2$  (10.4) and it was closely followed by  $AGL_3 \times CSR_2$  (9.7). Hybrids with average evaluation index value above 50 can be considered to possess greater economic value. The hybrid  $AGL_5 \times CSR_2$  exhibited maximum average evaluation index (68.52) followed by  $AGL_3 \times CSR_2$  (65.25).

Data revealed distinctive superiority of two hybrids  $AGL_3 \times CSR_2$  and  $AGL_5 \times CSR_2$ . No hybrid excelled in all the characters under study. Therefore, it is necessary to adopt reliable statistical methods for identification of promising hybrids giving weightage to different economic characters. In this direction, efforts have been made to identify promising silkworm hybrids utilizing multiple traits evaluation indices (Ramesh Babu *et al.*, 2002; Rao *et al.*, 2001, 2004; Lakshmi and Chandrashekharaiah, 2007). Hence in the present study, the indices obtained from the multiple traits evaluation method and subordinate function index were used and the two hybrids  $AGL_5 \times CSR_2$  and  $AGL_3 \times CSR_2$  were selected for field testing.

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### REFERENCES

- Gower, J. C. (1971) A general co-efficient of similarity and some of its properties. *Biometrics* **27**: 857–871.
- Lakshmi, H. and Chandrashekharaiah, (2007) Identification of breeding resource material for the development of thermo-tolerant breeds of silkworm, *Bombyx mori L. Journal of Experimental Zoology India* 10: 55–63.
- Mano, Y., Nirmal Kumar, S., Basavaraja, H. K., Mal Reddy, N. and Datta, R. K. (1993) A new method to select promising silkworm breeds/combinations. *Indian Silk* 31: 53.
- Narayanaswamy, T. K., Govindan, R. and Ananthanarayana, S. R. (2002) Selection of multivoltine × bivoltine cross breeds of silkworm, *Bombyx mori* L. through evaluation indices. *Indian Journal of Sericulture* **41**: 176–178.
- Ramesh Babu, M., Chandrashekharaiah., Lakshmi, H. and Prasad, J. (2002) Multiple traits evaluation of bivoltine hybrids of silkworm, *Bombyx mori* L. *International Journal of Industrial Entomology* 5: 37–44.
- Rao, C. G. P., Chandrashekharaiah., Basha, K. I., Seshagiri, S. V., Ramesh, C. and Nararaju, H. (2004) Idnetification of superior polyvoltine hybrids (Polyvoltine × bivoltine) of silkworm, Bombyx mori L. International Journal of Industrial Entomology 8: 43–49.
- Rao, C. G. P., Seshagiri, S. V., Ramesh, C., Basha, K., Ibrahim, H., Nagaraju, and Chandrashekaraiah, (2006) Evaluation of genetic potential of the polyvoltine silkworm (*Bombyx mori* L.) germplasm and identification of parents for breeding prograame. *Journal of Zhe-jiang University of Sciences* 7: 215–220.

Rao, P. S., Ravindra, S., Kalpana, G. V., Naik, V. N., Basavaraja, H. K., Rama Swamy, G. N. and Datta, R. K. (2001) Evaluation and identification of promising bivolitine hybrids of silkworm, *Bombyx mori* L. for tropics. *International Journal of Industrial Entomology* 3: 31–35.

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# Effect of feeding larvae of *Helicoverpa armigera* (Hubner) on Chickpea (*Cicer arietinum* L.) treated with chemical and organic fertilizers

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ABSTRACT: A field experiment was conducted to assess the effect of organic and chemical fertilizers used in cultivating chickpea (Cicer arietinum L.) on its major pest Helicoverpa armigera (Hubner). The sixth instar larvae were fed in the laboratory with leaves of the plants raised in the field experiment laid out in the research centre of the Narendra Dev University, Faizabad. The results showed that the application of NPK and farm yard manure increased the length and protein content of the larvae significantly when compared to the treatments with vermicompost as well as rhizobium for seed dressing and control. The weight of the larvae, and essential as well as non-essential aminoacid content of the larvae did not vary significantly from control. Rhizobium treated plants caused exceptionally low levels of free amino acids in the larvae. Significant differences were not seen in the effects of NPK and farm yard manure application. © 2008 Association for Advancement of Entomology

KEYWORDS: Helicoverpa armigera. effect of chemical/organic fertilizers, Cicer arietinum

Application of nitrogenous fertilizers in chickpea had been reported to enhance the development of larva and pupa of *Helicoverpa armigera* L., a regular pest on this crop. Viswanath (2005) observed that organic crops show higher tolerance to insect pests due to thicker cell wall and lower levels of free amino acids in the plants. Pest control through host nutrition manipulations has become a promising approach in recent IPM strategies (Giri *et al.*, 2004; Geire, 1986). In this context the effect of rearing larvae of *H. armigera* on chickpea grown under field conditions in Faizabad, giving chemical and organic fertilizers, was studied in the laboratory at Narendra Deva University of Agriculture and Technology, Faizabad U.P., India. The results are reported in this paper.

The field experiment was laid out adopting a completely randomized block design with a plot size of  $3 \times 2$  m. Udai variety of Chickpea (*C. arietinum*) was sown in the plots with the following fertilizer treatments applied as basal dose: (1) NPK @ 20:60:40 kg/ha, (2) Farm yard manure @ 8 t/ha, (3) vermicompost @ 37.5 q/ha.

(4) seed inoculation with *Rhizobium* culture @ 20mg/kg seed, and (5) control without any fertilizers.

H. armigera was reared in the laboratory using chick pea leaves collected from field. Leaves from the field experiment were collected plot wise and brought to the laboratory. Leaves from each plot were transferred to adequate number of Petri dishes (9 cm dia). First instar larvae of H. armigera, collected from the lab culture were individually transferred to the Petri dish. The larva in each dish was continuously supplied with leaves collected from the same plot. Adequate replications were maintained for each treatment. When the larvae reached sixth instar stage they were collected separately from each replication and their length was measured under a microscope. The weight of each larva was assessed using an electronic balance. Protein content of larva in each treatment was estimated following the methods of Lowry et al. (1951). Free amino acids and amides were estimated following the methods of Consden et al. (1944), Patridge (1948) and Bates et al. (1973). The data collected were subjected to statistical analysis and the results are presented in Table 1.

The data showed that the least length of the larva was in control (26.35 mm) and the highest length was in NPK treatment (29.90 mm) and the latter came on par with FYM (28.60 mm). The lengths of larvae in treatments with vermicompost (27.20 mm) and *Rhizobium* (27.60) were on par and significantly lower than those in FYM, but higher than that of control. Regarding the mean weights of the larvae in different treatments vermicompost treatment gave rise to larvae weighing significantly less (0.188 g) than the larvae in control (0.237 g) while the weight of larvae in remaining treatments (0.218 to 0.257 g) were on par with that of control.

Among the biochemical components of H. armigera larvae, protein content was seen significantly higher and on par with NPK, FYM and *Rhizobium* treatments (2427, 2240 and 2461  $\mu$ g/100 mg tissue) than in treatments with vermicompost and control, the latter two also being on par.

Seven essential aminoacids, leucine, isoleucine, valine, lysine, histidine, arginine, threonine, were identified in NPK. FYM and *Rhizobium* treatments and they are on par on weight basis. Of the above amino acids, lysine and histidine were lacking in vermicompost treatment while arginine and threonine were absent in control. The content of essential aminoacids was significantly higher in FYM and was very low in *Rhizobium* treatment compared to control. The remaining treatments were on par among themselves and also with control.

Results thus showed that the lengths of the larvae fed with leaves collected from chickpea plants treated with chemical fertilizers and farm yard manure were significantly higher than those obtained in other treatments. Vermicompost and rhizobium treatments also produced larvae longer than those in control and those treatments were on par but significantly less than NPK & FYM. With reference to the larval weight all the treatments came on par with control except vermicompost in which the larval weight was only 79% of the larval weight in control. Zing et al. (1982) observed that the application of nitrogen fertilizers in cotton plants caused an increase in the weight of *H. armigera* larvae. Purohit and Deshpande (1991) observed

TABLE 1. Effect of host nutrition on the length, weight, protein and amino acid content of sixth instar larvae of H. armigera.

Treat- ments	Length (mm)	Weight (gm)	Length Weight Protein (mm) (gm) (µg/100) mg									Amino acids 1/g/100 mg fresh weight	001/g//st	mg fresh	weight								
			fresh weight				Ewential									Non essential	ential						
				Leuk	Valine	Lys&	Arginine	Leu & Valine Lys & Arginine Threonine Total Total	Total	Total	n	Glycine Glut	Glut	Asp	Glut	Asp Glut y Amino fi alanine Tyrosine Aspara. Cystine Proline Total Total	alanine i	Tynasine	Aspara-	Cystine	Proline	Total	Tale
				Iso feur		Hist			lw.	Nos /	Vanine	Nos Alamine & Serine amic acid artic acid amine butyric	mic acid	artic acid	amine	butyric			gines			N.	Š.
NPK	29.90	0.257	2426	98.31	98.31 44.24	22.47	11.93	29,49	206.44 7.0		969	13.34	,	9.83	17.55	23.17	,	44.94	7.02	0.32	2.70	2.70 189.47 10	2
FYM	28 60	0.235	2240	100,001	99.19	114.16	(7.50)	28.33	321,65 7.0	7.0	5 19	71.66	ì	17.5	10.83	7.50	×.	21.66	10.0	3.30	6.67	236.62 10	0
Vernoi-	27.20	0.188	1527	89.70	0.98		11.01	10.11	213.94 5.11		75.0	13.97		12.5	22.0		47.05	15.20	,	21.99	× 2	280.10 9	0
compost																							
Rhizobium 27.60		0.218	2461	12.16	12.16 6.25	10.78	13.05	7.19	49,43 7.0		16%	,	7.74	3.87	,	,	,	2.26	0.60	1.54	13.54	15 37 7	į-
culture																							
Control	26.75	0.237	1385	76.12	56.08	96.15	ï	ì	228.35 5.0		55.28	41.40	,	17.6	20.03	41.66	28.84	44.07	,	,	5.78	254.68 9	2
69	0.40	0.023	325.04	10.4	5.3	6.0	132	2.13	24.1 0.71		7,49	3.93	0.22	1.89	2.46	2.07	1.87	3.14	0.42	1.88	0.91	25.37 1.09	1.0
27. 572																							

that the addition of nitrogenous fertilizers in host plants enhanced the food intake and consequently the larval weight. With reference to the length, the larvae of *H. armigera* in NPK and FYM treatment came on par and higher with consequent reduction in the tolerance/resistance in host plants. Advantage of using organic fertilizers over NPK (Viswanath, 2005) was not evident in the data of the experiment. The variation between the results of this experiment and earlier reports may be attributed to the soil plant interaction, or plant environment interaction in the field situations in Faizabad.

Protein content of the insects grown on hosts treated with NPK and FYM came on par and significantly higher than in control but it came on par with rhizobium treatment also, a treatment which did not cause higher growth of the larvae. Highest numbers of essential and non-essential amino acids were detected in NPK and FYM treatments as well as rhizobium treatments. But on the basis of the weight of the amino acids all the treatments came on par with control except rhizobium in which a very low content was observed. Thus the findings did not agree with the earlier reports that fertilizer application increases the weight (Purohit and Deshpande, 1991; Zing et al., 1982) protein (Haunerland et al., 1990) and free amino acids (Hedge, 2005). Adverse effects of inorganic fertilizers on the pest (Viswanath, 2005) also was not evident in the results of this experiment since FYM and NPK came on par throughout. The weight of essential and non-essential aminoacids observed in rhizobim treatment compared to control, was remarkably low and it may be attributed to some inhibitory effects. Lower length of the larvae in vermicompost and rhizobium treatments can also be attributed partly to the absence of some essential aminoacids in the treatments as observed by Chapman (1998) and Singh and Singh (2007).

### REFERENCES

- Bates, L. S., Walderen, R. P. and Teare, L. D. (1973) Rapid determination of free proline for water stress studies. *Plant Science* 39: 205–207.
- Chapman, R. F. (1998) Nutrition. In: *Insects Structure and Function*, Cambridge University Press, 69-91.
- Consden, R., Gordon, A. M. and Martin, A. J. P. (1944) Quantitative analysis of protein, a partition chromatographic method using paper. *Biochemistry Journal* 38: 224.
- Geire, P. W. (1986) Management of insect pest. Annals of Review Entomology 11: 471.
- Giri, A. P., Chougule, N. P., Telang, M. A. and Gupta, V. S. (2004) Engineering insect tolerant plant using plant defensive proteinase inhibitors. *Recent Research Development Phytochemistry* 8: 117-137.
- Haunerland, N. H., Nair, K. K. and Bowers, W. S. (1990) Fat body heterogeneity during developments of *Heliothis zea*. *Insect Biochemistry* 20: 829–837.
- Hegde, D. M. (2005) Eco-friendly agronomic practices for pest management, Winter School on Bio-intensive Integrated Management of Insect Pests and Disease of Crop, 15–18.
- Lowry, O. H., Rosebrough, H. J., Farr, A. L. and Randall, R. J. (1951) Protein measurement with folin phenol reagent. *Journal of Biological Chemistry* **193**: 265–275.
- Partridge, S. M. (1948) Filter paper partition chromatography of sugar: general description and application to the qualitative analysis of sugars in apple juice, egg white and foetal brood of sheep. *Biochemistry Journal* 42: 238–248.
- Purohit, M. S. and Deshpande, A. D. (1991) Effect of nitrogenous fertilizers of host plants on some growth parameters of *H. armigera* (Hub.). *Entomon* 16(2): 15–154.

- Singh, B. K. and Singh, R. P. (2007) Impact of feeding of chickpea (*Cicer arientinum* L.) raised by application of organic nutrients on length, weight, amino acids and protein contents of larvae of *H. armigera* (Hubner). In: *Abstracts Proceedings of National Seminar on Molecular Approaches for Crop Improvement During 7–8 February, 2007 at N.D.U.A.T. Kumarganj*, Faizabad, U.P. India.
- Viswanath, B. N. (2005) Management of pest and disease in erganic farming. In: Winter School on Bio-intensive Integrated Management of Insect Pests and Disease of Crop, 34-41.
- Zing, Y. L., Gong, P. Y., Jiang, L. R. and Zhang, M. L. (1982) Effect of nitrogen fertilizer application on cotton plant and the bollworm. *Acta-Entomologica-Sinica* **25(1)**: 16–23.

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# A new host plant record for *Oberea artocarpi* Gardner (Coleoptera: Cerambycidae)

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ABSTRACT: Ficus callosa Willd. (Moraceae) is reported as a new host plant of Oberea artocarpi Gardner (Coleoptera: Cerambycidae). © 2008 Association for Advancement of Entomology

KEYWORDS: Host plant, Ficus callosa, Oberea artocarpi, Kerala, India

The long horn beetle *Oberea artocarpi* Gardner is a minor pest of mulberry in south India. Its univoltine life cycle and nature of damage on mulberry were reported from Kerala (Prathapan 1995). The known host plants of *O. artocarpi* are *Artocarpus heterophyllus* Lam. (= *A. integrifolia* L.) (Gardner, 1941; Bhasin and Roonwal, 1954; Duffy, 1968) and *Morus alba* L. (Prathapan, 1995) (both Moraceae).

Infestation of *O. artocarpi* on young plants of *Ficus callosa* Willd. (Moraceae) was observed in a homestead at Koothattukulam. Kerala during 2007. Girdling of actively growing branches of about 8–9 mm diameter followed by die-back with ventilation holes is the typical symptom of infestation. An infested branch was collected on 6 November, 2007 and kept in the laboratory at College of Agriculture. Vellayani. Adult emerged on 25 March, 2008 through an oval exit hole of about 4.7 mm length and 4.0 mm width. Emergence of the adult coincided with the heavy pre-monsoon showers. *F. callosa* is a fast growing soft wood tree of little economic importance, except that its wood is used in the match industry. *O. artocarpi* appears to be specific to Moraceae utilizing different genera of the family as its host plants. This is the first report of *O. artocarpi* on *F. callosa*. The plant vouchers are deposited in the Calicut University Herbarium (Accession Nos 6323, 6324) and the insect specimen will be deposited in the National Pusa Collection. Division of Entomology, Indian Agricultural Research Institute. New Delhi.

### **ACKNOWLEDGEMENT**

F. callosa was identified by Dr A. K. Pradeep. Calicut University Herbarium.

### REFERENCES

- Bhasin, G. D. and Roonwal, M. D. (1954) A list of insect pests of forest plants in India and the Adjacent Countries (Arranged Alphabetically According to the Plant Genera and Species, For the Use of Forest Officers). Part 2. List of insect pests of plant genera 'A' (Aberia to Azima). Indian Forest Bulletin (New Series) Entomology 171(1): 5–91.
- Duffy, E. A. J. (1968) A Monograph of the Immature Stages of Oriental Timber Beetles (Cerambycidae). Publication No. 667, The British Museum (Natural History, 434.
- Gardner, J. C. M. (1941) Two new species of *Oberea* from Madras (Coleoptera: Cerambycidae). *Records of the Indian Museum* 43: 257–258.
- Prathapan, K. D. (1995) First Report of the incidence of *Oberea artocarpi* Gardner (Cerambycidae: Coleoptera) on mulberry. *Entomon* **20**(1): 67–69.

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Jalaja M. Muraleedharan D. and Prabhu V. K. K. (1973) Effect of extirpation of median neurosecretory cells on reproduction in the female red cotton bug, *Dysdercus cingulatus*. Journal of Insect Physiology 19(1): 29–36

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### **AUTHOR INDEX**

Ali, Haidar, 113 Ansari, M. S., 113

Cherian, P. T., 129

Ganga Visalakshy, P. N., 139 Gangopadhyay, D., 147 Geetha, P. R., 107 Gupta, Salil K., 119

Hasan, Wajid, 113

Jacob, Sosamma, 101

Karthikeyan, K., 101

Nirupama, R., 147

Prathapan, K. D., 157

Rana, Namrata, 135 Roy, Indranil, 119

Saha, Goutam K., 119 Saxena, Nishi, 135 Saxena, P. N., 135 Sharma, Harendra N., 135 Singh, B. K., 151 Singh, Jasvir, 91 Singh, Kan, 143 Singh, R. P., 151 Singh, Ravindra, 147 Sreelatha, K. B., 107 Swaminathan, R., 143

Tilak, Jyothi, 129